

Access to 2,6-Disubstituted Piperidines: Control of the Diastereoselectivity, Scope, and Limitations. Applications to the Stereoselective Synthesis of (–)-Solenopsine A and Alkaloid (+)-241D

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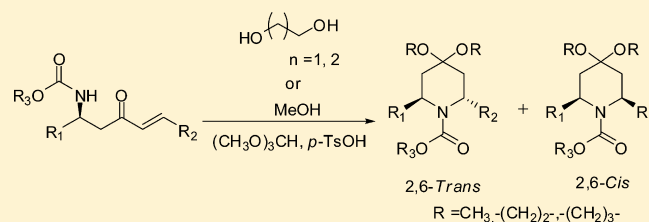
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S Supporting Information

ABSTRACT: Scope and limitations in the diastereoselective preparation of 2,6-*cis* or 2,6-*trans* disubstituted piperidines are described, through intramolecular reaction of chiral β' -carbamate- α,β -unsaturated ketone. This methodology has been applied to the total synthesis of a few well chosen examples, such as (–)-solenopsine A and alkaloid (+)-241D.



INTRODUCTION

Substituted piperidines and their analogues are key structural units in numerous naturally occurring alkaloids and in a number of successful pharmaceutical compounds¹ For this reason, a number of methodologies for the elaboration of these structures have been described^{2–5} especially when stereogenic centers are involved. In particular, those possessing a chiral center at C-2 and/or C-6, stereoselectivity that is essential for the defined activity, have attracted much attention because they are one of the most common framework encountered in many interesting compounds that exhibit a broad range of biological activities. For example, (–)-solenopsin A and (–)-isosolenopsin A (active components of fire ant venom) are reported to possess a broad range of activities,⁶ alkaloid (+)-241D (isolated from methanolic skin extracts of Panamanian poison frogs *Dendrobates speciosus*) is active on nicotinic acetylcholine receptors,⁷ and (–)-lasubine II (extracted from plants of the *Lythraceae* family) has showed cytotoxic, hemolytic, necrotic, insecticidal, antibacterial, antifungal, and anti-HIV properties⁸ (Scheme 1). So, developing approaches to allow the stereoselective synthesis of 2,6-dialkylpiperidines is of great value.

For this purpose, many synthetic methods have been developed including Mannich-type reactions⁹ or ring-closing metathesis.¹⁰ In order to control the diastereoselectivity excess on the positions α and α' of the piperidine core, some of those routes have focused on the construction of the ring by C–N ring-closure bond formation,^{11–13} including reductive amination,¹⁴ intramolecular substitution,¹⁵ cyclization of sulfinimides on propargylic ether,¹⁶ intramolecular allylic substitution with 1,3-chirality transfer,¹⁷ iminium ion cyclization,¹⁸ [4 + 2] cycloaddition of aldimines,¹⁹ intramolecular aza-[2,3]-Wittig rearrangement,²⁰ catalyzed hydroamination²¹ or Michael

addition.^{13,22} Therefore, all of these methods show that there is always a considerable interest in developing stereoselective access to 2,6-dialkylpiperidines. However, even though *cis*-2,6-disubstituted piperidines are readily accessible, only a few methods have been devoted to the synthesis of *trans*-2,6-disubstituted isomers.^{23–30}

During the course of our recent studies on the asymmetric synthesis of 2,6-disubstituted piperidines by C–N bond formation, we have demonstrated that the Michael-type cyclization,³¹ using β' -carbamate- α,β -unsaturated ketone **1** as key precursor, induced systematically and predominantly the formation of a piperidine ring with the 2,6-*trans* configuration (Scheme 2). The relative stereochemistry was confirmed by further transformation of the *trans* derivative in known chiral compound **3**³⁰ with an 95% ee.

In order to establish this new approach as a general method for the preparation of chiral 2,6-disubstituted piperidines and to understand the requirements for the best selectivity, we have synthesized various β' -carbamate- α,β -unsaturated ketones and tested their cyclization reaction using different conditions.

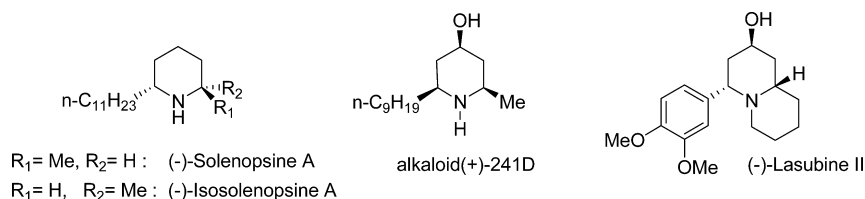
RESULTS AND DISCUSSION

General Synthesis of a Wide Range of β' -Carbamate- α,β -unsaturated Ketones. We have previously shown that the necessary β' -carbamate- α,β -unsaturated ketone **1** could be easily obtained from the corresponding α,β -unsaturated methylester in 6 steps with an overall yield of about 30%³¹ (Scheme 3).

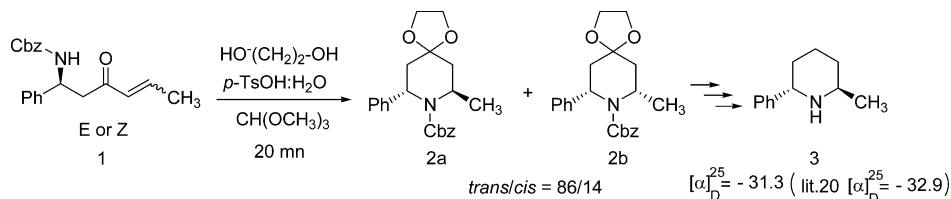
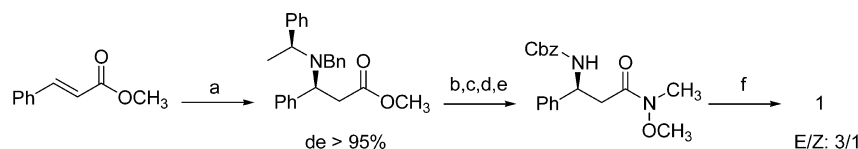
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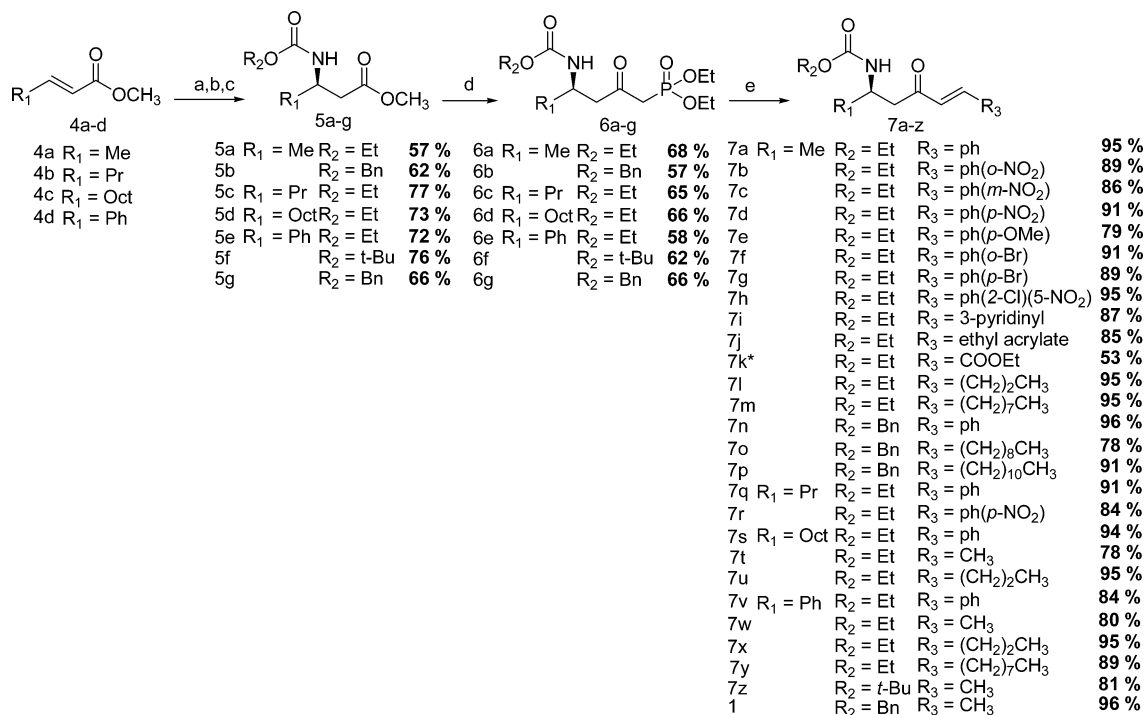
Scheme 1



Scheme 2

Scheme 3^a

^a(a) Davies amine, BuLi, THF, -78°C ; (b) H_2 , $\text{Pd}(\text{OH})_2/\text{C}$, MeOH; (c) Na_2CO_3 , CbzCl, $\text{CH}_2\text{Cl}_2/\text{H}_2\text{O}$; (d) NaOH 1 N, MeOH; (e) CDI, (MeO)MeNH.HCl; (f) Mg, 1-bromo-2-propene, THF, 0°C .

Scheme 4^a

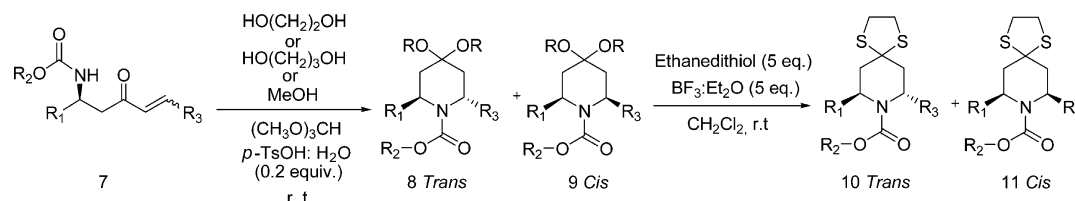
^a(a) Davies amine, BuLi, THF, -78°C ; (b) H_2 , $\text{Pd}(\text{OH})_2/\text{C}$, MeOH; (c) Na_2CO_3 , $\text{R}^2\text{CO}_2\text{Cl}$, $\text{CH}_2\text{Cl}_2/\text{H}_2\text{O}$; (d) BuLi, $(\text{EtO})_2\text{P}(\text{O})\text{Me}$, THF, -78°C ; (e) $\text{Ba}(\text{OH})_2$, THF/ H_2O (40:1), R^3CHO . In the case of enone 7k, we obtained a mixture of Z and E stereoisomers in a 60:40 ratio, respectively.

As Grignard's reagents do not allow the use of a wide range of functionalities, we have devised a general and simple method to access a variety of compounds of type 1 easily by using a more convenient way through a Wittig–Horner–Emmons³² reaction as the key step (Scheme 4). By this method, the

needed compounds were prepared in four steps from the corresponding α,β -unsaturated methylester according to described procedure.³³

Addition of enantiopure lithium *N*-benzyl-*N*- α -methylbenzylamide on α,β -unsaturated ester 4 following by hydrogenation

Scheme 5



to the corresponding primary amine and further protection as a carbamate gave the β -amino methylester **5a–g**. After purification, the ester function was transformed into the ketophosphonate **6a–g** by treatment with 2.5 equiv of diethyl lithiomethylphosphonate³⁴ in THF at -78 °C, in moderate yields. Over the years, many examples of base-promoted Wittig–Horner–Emmons reaction have been reported in scientific literature,^{35–37} and various combinations of bases and solvents (K_2CO_3/CH_3CN , DBU/THF, NaH/THF, $Et_3N/LiCl/CH_3CN$ or $Ba(OH)_2/(THF/H_2O)$, etc.) have been used. In our case, we have found that the use of 1.3 equiv of $Ba(OH)_2$ in biphasic medium THF/ H_2O (40:1) was the more general and convenient route to obtain compounds **7a–z** with good to excellent yield.

Scope and Limitations of the Michael-Type Cyclization. As mentioned previously,³¹ we have shown that compound **1** could be easily transformed diastereoselectively by intramolecular Michael-type reaction in 2,6-disubstituted-*N*-protected-4-ketal piperidine (**2a/2b**) as a mixture of *cis/trans* isomers in which the *trans* conformation represents the major compound (Scheme 2). We have also shown that the character *Z* or *E* of the geometry of the double bond in compound **1** did not have any influence on the diastereoselectivity of the cyclization reaction, as similar results have been obtained starting from either stereoisomer (*E*) or (*Z*) of **1** treated in the same optimized conditions (0.2 equiv of *p*-toluenesulphonic acid monohydrate, 5 equiv of ethylene glycol, 5 equiv of trimethyl orthoformate, which has been used here as solvent and as a water scavenger).

So, we decided to use this protocol for the cyclization of a range of dissymmetric (aliphatic/aromatic for R^1 and R^3) enecarbamates of type **7**, hoping to evaluate at first the influence of steric hindrance on the selectivity. For a better evaluation of the *cis/trans* ratio (1H NMR) the mixture of ketals **8** and **9** were directly converted into the more stable thioketals **10** and **11** by known procedure, using 1,2-ethane dithiol in the presence of boron trifluoride diethyl etherate, as it has been shown that this transformation induced no variation of the diastereoisomeric ratio (Scheme 5 and Table 1).

According to Table 1, for the defined conditions, the selectivity observed for the cyclization reaction is predominantly in favor of the *trans* isomer, which is the less stable conformation for a 2,6-disubstituted piperidine. This *de* is markedly dependent on a lot of factors, namely, the nature of the nitrogen protective group and also the nature of steric hindrance (R^1 , R^2 , and R^3) on compounds **7**. On the one hand, when R^1 and R^3 are fixed ($R^1 = Ph$, $R^3 = Me$; entries 1, 2, and 3), a strong steric hindrance around the nitrogen atom is necessary to induce a good diastereoselectivity. On the other hand, when $R^1 = Ph$ and R^3 is a much longer alkyl chain (propyl or nonyl; entry 7, 8, and 9), the ethyl carbamate function is sufficient to ensure predominantly the formation of the *trans* isomer, however with a small diastereoselectivity

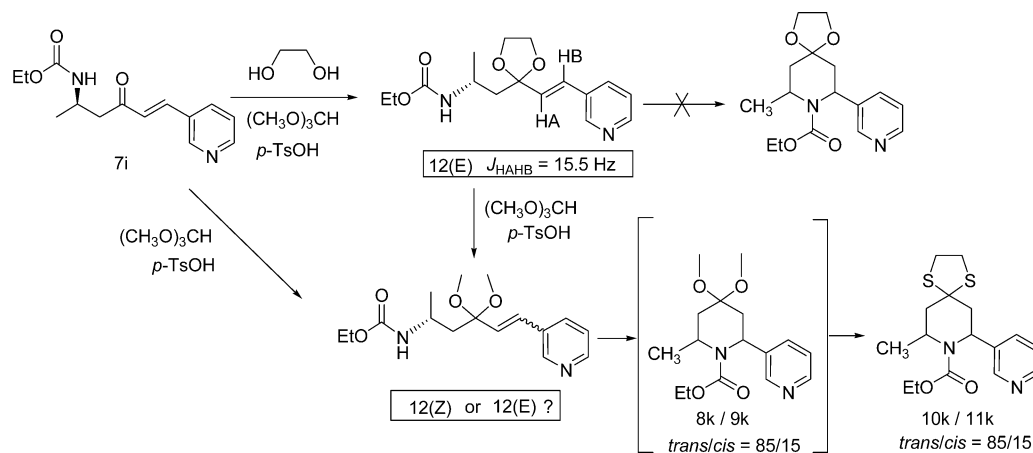
Table 1. Scope of Intramolecular Michael Reaction

Entry	Ketone 7	cyclization react. time (min)	alcohol	(<i>Trans/Cis</i>) 8/9 (%) ^a	(<i>Trans/Cis</i>) 10/11 [% ^b]
1		20 min	HO(CH ₂) ₂ OH	86/14	86/14 [68] ^c
2		30 min	HO(CH ₂) ₂ OH	83/17	83/17 [71]
3		20 min	HO(CH ₂) ₂ OH	44/56	44/56 [75]
4		20 min	HO(CH ₂) ₂ OH	51/49	51/49 [65]*
5		20 min	HO(CH ₂) ₂ OH	39/61	39/61 [73]
6		40 min	HO(CH ₂) ₂ OH	≥ 95/5	≥ 95/5 [60]
7		40 min	HO(CH ₂) ₂ OH	62/38	62/38 [67]
8		2 h	HO(CH ₂) ₂ OH	62/38	62/38 [65]
9		2 h	HO(CH ₂) ₃ OH	95/5	95/5 [67]
10		1 h	HO(CH ₂) ₂ OH	95/5	95/5 [67]
11		2 h	HO(CH ₂) ₂ OH	≥ 96/4	≥ 96/4 [65]
12		2 h	HO(CH ₂) ₂ OH	see text	
13		6 h	MeOH	85/15	85/15 [48]

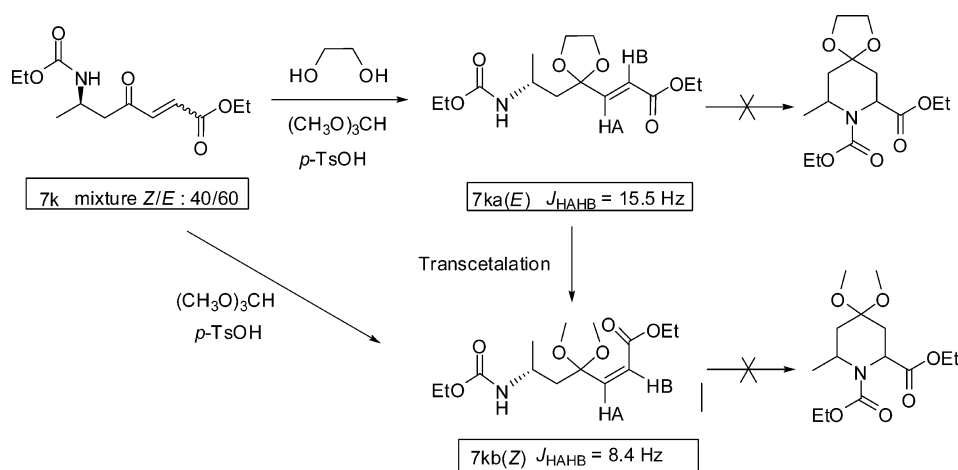
^aDiastereoisomeric ratio is determined by integration of characteristic protons of the piperidine ring in the crude H^1 NMR spectra (*trans* and *cis* isomers were determined according to their respective coupling constants). ^bIsolated yield of pure diastereoisomeric mixture after chromatography on silica gel. ^c**10a/11a** and **10d/11d** are enantiomers; for **10a/11a** (**10d/11d**) the benzyl carbamate was cleaved during the thioketalation process, leading to the free amine on the piperidine ring, see Experimental Section).

excess (*de* = 24%). Furthermore, changing ethylene glycol to 1,3-propane diol increases the *de* up to 90%, showing by this way the importance of the resulting keto protecting group. Permutation of R^1 and R^3 (entry 3 versus entry 5) does not show a significant influence on the diastereoselectivity unless the alkyl chain is much longer than a methyl group (entries 5, 6, and 10), or if R^1 is a phenyl group (entry 11), in which the observed *de* is again around 90%. Another observation is the dependency of the cyclization on the nature of the alcohol used. So, when $R^1 = Me$ and $R^3 = pyridine$ (**7i**), the formation of the

Scheme 6



Scheme 7



piperidine ring is not observed if ethylene glycol is used to form the ketal (entry 12). The only product that can be identified (by ^1H NMR spectroscopy) is the ketal **12**, in which the double bond is exclusively in a *E* conformation ($J = 15.5 \text{ Hz}$). Assuming that the relative hindrance between the dioxolane group and the pyridine was too high, we decided to realize the reaction with a less crowded acetal. To our delight, when compound **7i** was engaged (entry 13) in the presence of trimethyl orthoformate and *p*-toluenesulphonic acid, leading to the *in situ* formation of methanol, we could now isolate the corresponding piperidines (**10k/11k**) with a de of 70% in favor of the *trans* conformation (Scheme 6).

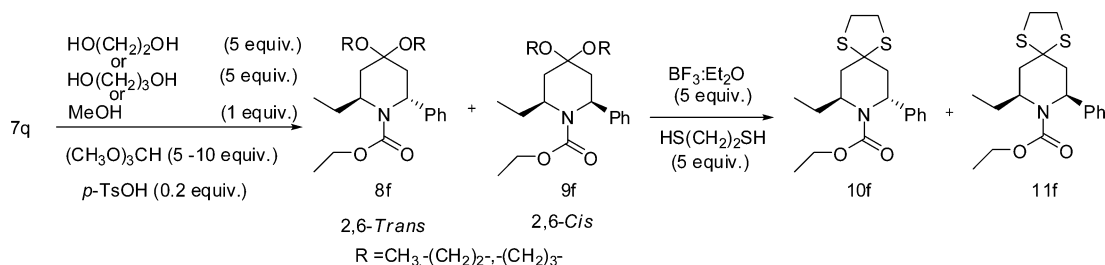
This last result confirmed the fact that the first step, for the elaboration of the piperidine ring, is the formation of the ketal on compounds **7** and thus can constitute the critical step for the diastereoselectivity of the reaction. Moreover, the formation of the ketal induces the *E* or *Z* configuration of the double bond, which is dependent on the nature of the ketal. This hypothesis based on the role of the geometry of the double bond of a Michael acceptor in the control of the diastereoselectivity, during the formation of 2,6-disubstituted piperidines, has already been put forward by Banwell and co-workers,³⁸ although in the case of an exocyclization process. They had demonstrated that the geometry of the double bond conducted to two different transition states, resulting in the formation of 2,6-*cis* or 2,6-*trans* piperidine. Thus, reducing this fact to our model, we suppose that the formation of the 2,6-*trans*

piperidine or 2,6-*cis* piperidine can be correlated with the geometry of the double bond of the crucial intermediate acetal.

In order to confirm this hypothesis we engaged the compound **7j** (*Z/E* conformers = 40/60) in reaction with trimethyl orthoformate, with and without ethylene glycol and in the presence of acid (Scheme 7). Compound **7j** was specially chosen for the strong steric hindrance that could be generated in the transition state. At this stage, no formation of a piperidine ring was expected, but rather the formation of the intermediate ketal form and the possibility to measure the corresponding coupling constant of both intermediates, to validate our hypothesis.

Then, it was possible to identify two α,β -unsaturated ketals **7ka(E)** and **7kb(Z)**, a dioxolane and a dimethylketal, respectively. The coupling constant values for the double bond in ^1H NMR spectroscopy, demonstrated the existence of two different stereoisomers depending on the ketal formed. The *E* configuration ($J_{\text{HAHB}} = 15.5 \text{ Hz}$) was observed for the cyclic ketal **7ka(E)**, whereas the *Z* ($J_{\text{HAHB}} = 8.4 \text{ Hz}$) was observed for the dimethyl ketal **7kb(Z)**. As for compound **7ka(E)**, if trimethyl orthoformate is added to the mixture, a transacetalation reaction is observed, leading to the formation of **7kb(Z)**. Thus, this result first of all confirmed the formation of the ketal as the first step of the transformation and, second, strongly suggested that the diastereoselectivity of the piperidine formed can be dependent on the configuration of the double bond of the intermediate ketal in the transition state. However

Table 2. Influence of Ketal Formation on the Stereoselectivity



entry	alcohol used	cyclization reaction time 8f/9f (min)	10f/11f (ratio ^a %) [yield ^b %]
1	HO-(CH ₂) ₂ -OH (7qR)	20	>95/5 [67]
2	HO-(CH ₂) ₂ -OH (7qS)	20	>95/5 [68]
3	HO-(CH ₂) ₃ -OH	15	87/13 [68]
4	MeOH	15	64/36 [68]

^aDiastereoisomeric ratio is determined by integration of characteristic protons of the piperidine ring in the crude ¹H NMR spectra (*trans* and *cis* isomers were determined according to their respective coupling constants). ^bIsolated yield of pure diastereoisomeric mixture after chromatography on silica gel.

at this stage we cannot connect the configuration of the double bond with the configuration of the piperidine formed. We can only assume the existence of two different transition states corresponding to the formation of two different acetals.

Therefore, to confirm the real role of this geometry on the resulting diastereoisomeric excess for the piperidine generated, we selected the compound **7q**, which gave a de ≥ 90% in favor of the *trans* isomer (Table 1, entry 6) and compared the importance of the nature of the alcohol on the result when methanol, ethylene glycol, or propan-1,3-diol is used in the cyclization process (Table 2); both enantiomers of **7q** were tested. The quantity of acid was fixed at 0.2 equiv, and all the crude mixtures **8f,9f** were directly converted into, respectively, the corresponding 4-thioacetal piperidine **10f** and **11f** (Table 2). The diastereomeric excess was as before, calculated according to the ¹H NMR values.

Cyclization was observed in all cases with a good overall yield, but a significant difference in the de was observed. The higher diastereoselectivity was obtained (Table 2, entry 1 and 2) when ethylene glycol is used to form the ketal, corresponding to the more overcrowded intermediate. On the contrary, lower diastereoselectivity is observed when methanol is used (Table 2, entry 4), as the dimethyl acetal gave a higher flexibility to the intermediate. Both enantiomers of **7q** gave the same result (Table 2, entries 1 and 2). If steric hindrance appeared here as the predominant factor for the stereoselectivity of the reaction, however, in all cases the *trans* isomer was obtained. So, in order to reinforce the existence of two different transition state according to the geometry of the double bond of the ketal, we envisaged that this critical step could be under a kinetic control. Compound **7q** was now engaged under two different experimental protocols: on the one hand with ethylene glycol and on the other hand with methanol, and for each case increasing quantities of acid, from catalytic to stoichiometric, were used (Table 3).

When ethylene glycol is used for the formation of the ketal, the quantity of acid does not affect the diastereoisomeric excess obtained for **10f/11f** (entries 1–4), and only the reaction time is reduced. After 24 h (entry 5), an epimerization through retro-Mannich or retro-Michael reaction is observed, the more stable *cis* isomer **11f** becoming now the major compound. On the contrary, when methanol is used (entries 6–10) to generate the ketal, there is a significant difference in the diastereoisomeric

Table 3. Kinetic Effect on the Stereoselectivity in the Cyclization Process

entry	alcohol (equiv)	$p\text{-TsOH}/\text{H}_2\text{O}$ (equiv)	cyclization reaction time 8f/9f (min)	10f/11f (ratio ^a %)
1	HO-(CH ₂) ₂ -OH (5)	0.1	25	85/15
2	HO-(CH ₂) ₂ -OH (5)	0.2	20	85/15
3	HO-(CH ₂) ₂ -OH (5)	0.5	15	86/14
4	HO-(CH ₂) ₂ -OH (5)	1	10	87/13
5	HO-(CH ₂) ₂ -OH (5)	0.2	24 h	40/60
6	MeOH (1)	0.02	45	50/50
7	MeOH (1)	0.05	40	55/45
8	MeOH (1)	0.1	20	64/36
9	MeOH (0 or 1)	0.2	15	69/31
10	MeOH (0) ^b	1	10	72/28

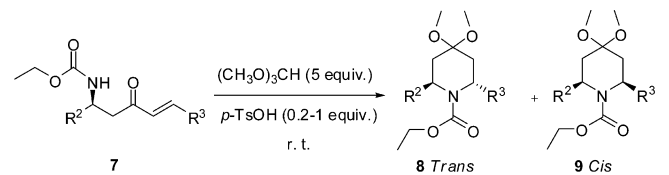
^aDiastereoisomeric ratio is determined by integration of characteristic protons of the piperidine ring in the crude ¹H NMR spectra (*trans* and *cis* isomers were determined according to their respective coupling constants). ^bWhen the concentration of acid is more than 0.2 equiv, the degradation of trimethyl orthoformate is sufficient to generate methanol *in situ*.

meric excess depending on the quantity of catalyst used, and this evolution is in agreement with a kinetic effect.

To evaluate the importance of this kinetic effect in the transition state, compared to the steric effect, we substituted the aromatic ring of **7a** and **7q** (respectively table 3, entry 5, 39% of *trans* isomer and entry 6, ≥ 95% of *trans* isomer, with the use of ethylene glycol and 0.2% acid) with various ERG or EWG groups in the *ortho*, *meta*, or *para* position of the aromatic ring, and we used the methanol generated by the trimethyl orthoformate for the formation of the ketal. Results on the selectivity so obtained are reported in Table 4.

Strong EWG located in *para* position on the aromatic ring for **7r** and **7d** (Table 4, entries 1 and 2) or placed on a conjugated system **7j** (Table 4, entry 9) led predominantly and respectively to 83%, 89%, or 90% in favor to the *trans* isomer, but no significant influence on the selectivity can be related to the quantity of acid used. When the EWG is in the *meta* position, (compound **7c**, Table 4, entry 3), a deactivating position, the opposite diastereoselectivity (40/60) is observed and a longer reaction time is required. Concerning the *ortho* position (compound **7b**, Table 4, entry 4), the EWG effect is

Table 4. Electronic Effects



Entry	Ketone 7	Reaction time (min)	<i>p</i> -TsOH/H ₂ O (x equiv.)	8/9 (%) ^a [yield %] ^b
1		(90min)	(0.2 equiv.)	83/17 [71] (8l/9l)
2		(120min)	(0.2 equiv.)	89/11 [80] (8m/9m)
3		(60min)	(1 equiv.)	40/60 [71] (8n/9n)
4		(60min)	(1 equiv.)	60/40 [67] (8o/9o)
5		(45 min)	(1 equiv.)	33/67 [73] (8p/9p)
6		(60min)	(1 equiv.)	29/71 [75] (8q/9q)
7		(180min)	(1 equiv.)	56/43 [76] (8r/9r)
8				decomposition
9		(240min)	(0.2 equiv.)	90/10 [93] (8s/9s)

^aDiastereoisomeric ratio is determined by integration of characteristic protons of the piperidine ring in the crude ¹H NMR spectra (*trans* and *cis* isomers were determined according to their respective coupling constants). ^bIsolated yield of pure diastereoisomeric mixture after chromatography on silica gel.

counterbalanced by steric hindrance in the transition state, and therefore a small diastereoisomeric excess (60/40) is obtained. As expected, the presence of an ERG group in the *ortho* or *para* position (Table 4, entries 5 and 6) led predominantly to the *cis* derivative (30/70). However, if this effect is too strong, decomposition of the starting material is observed (Table 4, entry 8). Thus, when compounds 7 have an aromatic or a conjugated system as a substituent on the double bond, a lot of parameters (steric hindrance, angle pressure, kinetic effect and electronic effect) has to be considered to access a high selectivity, and this selectivity is in favor of the *trans* isomer of the piperidine.

So, in order to validate the existence of two transition states, according to the alcohol used in the cyclization step, we carried out the reaction with four representative ketones 7k,l,t,u in which only a steric hindrance was induced by the size of an alkyl chain. As usual, we measured the outcomes observed for the diastereoselectivity when a cyclic ketal or a dimethyl ketal formation was involved. Here too, we converted all the crude

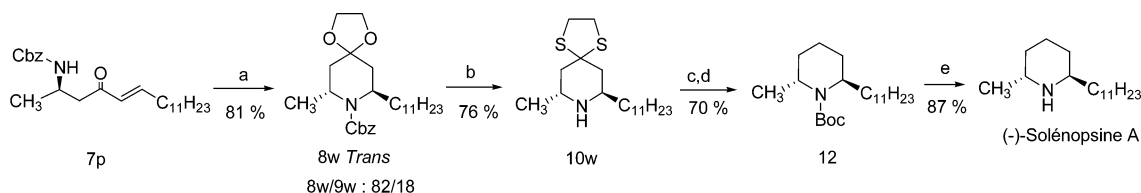
mixture 8 and 9 or 8' and 9' directly into the corresponding 4-thioetheral piperidines 10 and 11 (Table 5).

According to the results obtained (Table 5) and the stereoselectivity observed, it becomes evident now to affirm the formation of two different transition states in the formation of the piperidine ring. Both transition states are strongly dependent on the alcohol used to generate the ketal on the α,β -unsaturated ketone. As we have mentioned in Table 3, there are two behaviors according the nature of the ketal group. When dioxolane is used, the *trans* isomer is the major product formed, at ~80%, with a catalytic amount of acid (Table 5, entries 1, 4, 5, 7). However when a stoichiometric quantity of acid is used, a fast epimerization process occurs (retro Mannich or Michael reaction), and consequently a higher formation of the thermodynamic specie, namely, the 2,6-*cis* piperidine, is observed, even if the *trans* isomer was the first formed. In contrast, as observed in table 3, there is no kinetic effect on the cyclization when methanol is used. In this case, the *cis* isomer was the major product, ~80% of the reaction; whatever the quantity of acid engaged (Table 5, entries 3, 6, 8), the

Table 5. Michael Cyclization of Enones 7 Bearing Aliphatic Substituents

Entry	Ketone 7	Cyclisation (React. time)	Alcohol	(<i>Trans/Cis</i>) 10/11 (% ^a) [yield %]
		<i>p</i> -TsOH [x equiv.]		
1		40 min [0.2]	HO(CH ₂) ₂ OH	80/20 [73] (10v/11v)
2		1 h [0.2]	HO(CH ₂) ₂ OH	80/20 [62] (10t/11t)
3		1 h [0.2]	MeOH ^c	20/80 [66] (10t/11t)
4		2 h [0.2]	HO(CH ₂) ₂ OH	82/18 [68] (10s/11s)
5		0.5 h [1]	HO(CH ₂) ₂ OH	65/35 [70] (10s/11s)
6		0.5 h [0.2]	MeOH ^c	20/80 [71] (10s/11s)
7		0.5 h [1]	MeOH ^c	20/80 [69] (10s/11s)
8		2 h [0.2]	HO(CH ₂) ₂ OH	83/17 [70] (10u/11u)

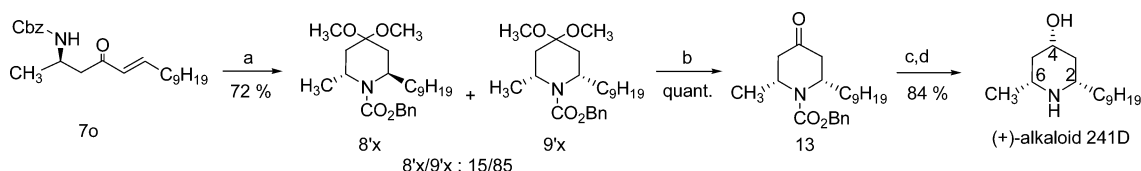
^aDiastereoisomeric ratio is determined by integration of characteristic protons of the piperidine ring in the crude ¹H NMR spectra (*trans* and *cis* isomers were determined according to their respective coupling constants). ^bIsolated yield of pure compounds after chromatography on silica gel. ^cMeOH was generated *in situ* by decomposition of trimethyl orthoformate.

Scheme 8^a

^a(a) OH(CH₂)₂OH, (CH₃O)₃CH, *p*-TsOH; (b) SH(CH₂)₂SH, BF₃·Et₂O, CH₂Cl₂; (c) (Boc)₂O, DMAP, CH₂Cl₂; (d) W-2 Raney nickel, EtOH, reflux; (e) TFA, CH₂Cl₂.

thermodynamic product was formed predominantly. Then, as for Banwell's studies, our results are in agreement with the formation of two possible conformations of the conformation of the double bond *before* nucleophilic attack of the carbamate, leading to the formation of the piperidine. At this stage, after examination of all potential parameters that can

interfere with diastereoselectivity, it becomes easy to prepare either the *trans* or the *cis* isomer by a wise choice of experimental conditions. In order to demonstrate this, we applied these protocols to the synthesis of (–)-solenopsin-A^{39,40} and (+)-alkaloid 241D^{41–43} (Schemes 8 and 9). (–)-Solenopsin A can be rapidly prepared from compound

Scheme 9^a

^a(a) $(\text{CH}_3\text{O})_3\text{CH}$, pTsOH; (b) TFA/ H_2O , CH_2Cl_2 ; (c) Pd/C (5%), MeOH, H_2 , 1 atm; (d) NaBH_4 , MeOH.

7p by using for the cyclization step ethylene glycol, which led to the *trans* isomer 8w as a major product (82%, Scheme 8). After subsequent transformations that we have already described in a previous paper⁴⁴ (thioacetalation 10w, Boc protection, desulfuration 12), (–)-solenopsin A was obtained in 10 steps, after regeneration of the free amine, from the α,β -ethylenic ester 4, with an overall yield of 9% with $[\alpha]_{\text{D}} = -1.21$ (*c* 0.94 MeOH, lit.³⁴ $[\alpha]_{\text{D}} = -1.30$ (*c* 1.30 MeOH)).

To reach (+)-alkaloid 241D, methanol was used for the cyclization, starting from compound 7o. The intermediates 8'x/9'x were obtained in a ratio of 15/85 in a favor of the *cis* isomer. After separation on deactivated silica, 9'x was keto-deprotected using a 40% aqueous trifluoroacetic acid solution at room temperature to give the corresponding piperidones 13 in very good yield. *N*-Deprotection of the piperidone 13, followed by reduction with NaBH_4 gave selectively (+)-alkaloid 241D in 9 steps from the α,β -ethylenic ester 4 with an overall yield of 16% with a de $\geq 95\%$ and an ee of 92% (Scheme 9).

CONCLUSION

In conclusion, we have described herein a methodology to prepare stereoselectively either 2,6-*cis* or 2,6-*trans* disubstituted piperidines. The efficiency of this methodology has been demonstrated through the asymmetric synthesis of (–)-solenopsin A and (+)-alkaloid 241D together with their respective isomer in C-6, demonstrating in this way that this strategy will be applied efficiently to the total synthesis of other piperidinic alkaloids exhibiting important biological interest.

EXPERIMENTAL SECTION

General. Organic solutions were dried over Na_2SO_4 and filtered. When anhydrous solvents were used, they were prepared as follows: tetrahydrofuran (THF) was distilled under N_2 from sodium benzophenone ketyl and used immediately; anhydrous acetonitrile was freshly distilled from CaH_2 . All ^1H NMR and ^{13}C spectra were measured in CDCl_3 or C_6D_6 and recorded on a Bruker 400 MHz (101 MHz for ^{13}C) spectrometer using TMS as the internal standard. Chemical shifts are expressed in ppm and *J* values are given in hertz. The following abbreviations are used: singlet (s), broad singlet (brs), doublet (d), doubled doublet (dd), triplet (t), multiplet (m). High resolution mass spectroscopy (HRMS, TOF) were carried out in electrospray mode. Monitoring of the reactions was performed using silica gel TLC plates. Spots were visualized by UV light at 254 nm. Flash chromatography columns were performed using silica gel 60 (70–230 mesh).

General Procedure for the Synthesis of β -Aminoesters 5. (R)-Methyl-3-(ethoxycarbonylamino)butanoate 5a. To a cold solution (0 °C) of (+)-(R)-*N*-benzyl-*N*- α -methyl benzylamine (23.0 mL, 110 mmol, 1.1 equiv) in dry THF (280 mL) was added slowly under argon *n*-butyl lithium (75.0 mL, 1.6 M in hexane, 120 mmol, 1.2 equiv). The resultant pink solution of lithium amide was stirred for 30 min at 0 °C and then cooled to –78 °C before dropwise addition of a solution of methyl crotonate (10.0 mL, 100 mmol, 1 equiv) in dry THF (100 mL). The mixture was stirred at –78 °C for 3.5 h. Then, a saturated aqueous solution of NH_4Cl (100 mL) was added slowly, and the

resulting solution was allowed to warm to room temperature. The solution was extracted twice with ethyl acetate. Combined organic extracts were dried over Na_2SO_4 , filtered, and evaporated. The crude product was added to a suspension of 10% Pd/C (5.00 g) in methanol (200 mL). The mixture was placed on a Parr apparatus and stirred under a hydrogen atmosphere (60 psi) for 4 days. The catalyst was then removed by filtration on Celite. The residue was concentrated *in vacuo* and dissolved in dichloromethane (200 mL) and water (200 mL). Then, sodium carbonate (42.4 g, 400 mmol, 4.0 equiv) and ethyl chloroformate (28.5 mL, 200 mmol, 2 equiv) were added dropwise. The resulting solution was stirred at room temperature for 3 h. The aqueous material was extracted with dichloromethane and the combined organic extracts were dried over Na_2SO_4 , filtered, and concentrated *in vacuo*. Purification by chromatography on silica gel (cyclohexane/EtOAc 9:1 to 5:5) afforded 5a as a yellow oil (10.8 g, 57% over 3 steps): $[\alpha]_{\text{D}} = -35.60$ (*c* 0.99, CHCl_3), lit.²⁷ $[\alpha]_{\text{D}} = -37.07$ (*c* 1.00, CHCl_3). ^1H NMR (400 MHz, CDCl_3) δ 5.03 (brs, 1H, NH), 4.03 (m, 3H), 3.62 (s, 3H), 2.46 (d, *J* = 6.9 Hz, 2H), 1.16 (t, *J* = 6.9 Hz, 3H), 1.15 (d, *J* = 6.6 Hz, 3H). Spectral data are identical with those reported.²⁷

(R)-Methyl-3-(ethoxycarbonylamino)butanoate 5a. (Starting from 0.100 mol of 4a) Yellow oil, 10.8 g, yield = 57 % Spectral data are identical with those reported:⁴⁵ $[\alpha]_{\text{D}} = -37.07$ (*c* 1, CHCl_3). ^1H NMR (400 MHz, CDCl_3) δ 5.03 (brs, 1H), 4.03 (m, 3H), 3.62 (s, 3H), 2.46 (d, *J* = 6.9 Hz, 2H), 1.16 (t, *J* = 6.9 Hz, 3H), 1.15 (d, *J* = 6.6 Hz, 3H).

(R)-Methyl-3-(benzyloxycarbonylamino)butanoate 5b. (Starting from 0.100 mol of 4a) Yellow oil, 15.6 g, yield = 62%. Spectral data are identical with those reported:⁴⁶ $[\alpha]_{\text{D}} = +16.9$ (*c* 1.4, CHCl_3). ^1H NMR (400 MHz, CDCl_3) δ 5.03 (brs, 1H), 4.03 (m, 3H), 3.62 (s, 3H), 2.46 (d, *J* = 6.9 Hz, 2H), 1.16 (t, *J* = 6.9 Hz, 3H), 1.15 (d, *J* = 6.6, 3H).

(R)-Methyl-3-(benzyloxycarbonylamino)butanoate 5c and (S)-Methyl-3-(benzyloxycarbonylamino)butanoate 5c'. (Starting from 0.100 mol of 4b) Yellow oil, 16.7 g, yield = 77%. *R* enantiomer: $[\alpha]_{\text{D}} = +41.5$ (*c* 1.03, CHCl_3), *S* enantiomer: $[\alpha]_{\text{D}} = -40.9$ (*c* 1.035, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 4.98 (brs, 1H), 4.02 (m, 2H), 3.90 (m, 1H), 3.61 (s, 3H), 2.49 (dd, *J* = 15.8, 4.8 Hz, 1H), 2.43 (dd, *J* = 15.8, 5.3 Hz, 1H), 1.48–1.22 (m, 2H), 1.16 (t, *J* = 7.0 Hz, 3H), 0.85 (t, *J* = 7.2 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.0, 156.0, 60.7, 51.6, 47.7, 38.9, 36.6, 19.3, 14.6, 13.8. HRMS-ESI (M + Na) *m/z* calcd for $\text{C}_{10}\text{H}_{19}\text{NO}_4\text{Na}$ 240.1212, found 240.1216.

(R)-Methyl-3-(ethoxycarbonylamino)undecanoate 5d. (Starting from 0.100 mol of 4c) Yellow oil, 20.9 g, yield = 73%. $[\alpha]_{\text{D}} = +29.2$ (*c* 1.05, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 5.07 (brs, 1H), 4.22–4.03 (m, 2H), 3.98 (m, 1H), 3.69 (s, 3H), 2.57 (dd, *J* = 15.1, 4.6 Hz, 1H), 2.51 (dd, *J* = 15.1, 5.1 Hz, 1H), 1.55–1.45 (m, 2H), 1.41–1.10 (m, 15H), 0.89 (t, *J* = 6.9 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.1, 156.1, 60.7, 51.5, 48.0, 38.9, 34.4, 31.8, 29.4, 29.3, 29.2, 26.1, 22.6, 14.6, 14.1; HRMS-ESI (M + Na) *m/z* calcd for $\text{C}_{15}\text{H}_{29}\text{NO}_4\text{Na}$ 310.1994, found 310.1996.

(S)-Methyl-3-(ethoxycarbonylamino)-3-phenylpropanoate 5e. (Starting from 0.100 mol of 4d) Yellow oil, 18.0 g, yield = 72%. $[\alpha]_{\text{D}} = -9.7$ (*c* 0.99, CHCl_3). ^1H NMR (400 MHz, CDCl_3) δ 7.36–7.25 (m, 5H), 5.75 (brs, 1H), 5.17 (m, 1H), 4.11 (q, *J* = 7.0 Hz, 2H), 3.62 (s, 3H), 2.91 (dd, *J* = 15.5, 6.0 Hz, 1H), 2.84 (dd, *J* = 15.5, 5.9 Hz, 1H), 1.23 (t, *J* = 7.0, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 171.3, 155.8, 140.9, 128.6, 127.6, 126.2, 61.0, 51.8, 51.7, 40.5, 14.6; HRMS-ESI (M + Na) *m/z* calcd for $\text{C}_{13}\text{H}_{17}\text{NO}_4\text{Na}$ 274.1055, found 274.1069.

(*S*)-Methyl-3-(*tert*-butoxycarbonylamino)-3-phenylpropanoate **5f**. (Starting from 0.100 mol of **4d**) Yellow oil, 21.2 g, yield = 76%. Spectral data are identical with those reported.⁴⁷

(*S*)-Methyl-3-(benzyloxycarbonylamino)-3-phenylpropanoate **5g**. (Starting from 0.100 mol of **4d**) Yellow oil, 20.7 g, yield = 66%. Spectral data are identical with those reported⁵²: $[\alpha]_D = -16.1$ (c 0.97, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.22 (m, 10 H), 5.73 (brs, 1H), 5.09 (m, 1H), 5.01 (d, *J* = 12.3 Hz, 1H), 4.97 (d, *J* = 12.3 Hz, 1H), 3.50 (s, 3H), 2.81 (dd, *J* = 15.3, 5.0 Hz, 1H), 2.74 (dd, *J* = 15.3, 5.7 Hz, 1H).

General Procedure for the Synthesis of Ketophosphonates

6. (R)-Ethyl 5-(Diethoxyphosphoryl)-4-oxopentan-2-ylcarbamate 6a. To a solution of diethyl methylphosphonate (5.8 mL, 39.7 mmol, 2.5 equiv) in anhydrous THF (15 mL) at -78 °C was added dropwise *n*-butyl lithium (24.8 mL, 1.6 M in hexane, 39.7 mmol, 2.5 equiv). After 20 min at -78 °C, a solution of **5a** (3 g, 15.9 mmol, 1 equiv) in anhydrous THF (15 mL) was added dropwise. After addition, the temperature of the reaction was kept at -78 °C for 30 min, then allowed to reach 0 °C in 1 h, quenched with a solution of ammonium chloride, and extracted twice with ethyl acetate. After drying over Na₂SO₄ and concentration under vacuum, the crude oil was first distilled at low pressure to remove excess diethyl methylphosphonate, and the residue purified by flash chromatography (eluent, cyclohexane/EtOAc 2:1 to EtOAc) afforded compound **6a** as a yellow oil (3.3 g, 68% yield): $[\alpha]_D = +33.60$ (c 1.17, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 5.03 (brs, 1H), 4.16–3.94 (m, 7H), 3.08 (dd, *J* = 23.0, 14.0 Hz, 1H), 2.99 (dd, *J* = 22.6, 14.0 Hz, 1H), 2.84 (dd, *J* = 17.1, 6.0 Hz, 1H), 2.71 (dd, *J* = 17.1, 5.7 Hz, 1H), 1.33–1.21 (m, 6H), 1.15–1.20 (m, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 200.6, 155.8, 62.6 (d, *J* = 6.6 Hz), 62.5 (d, *J* = 6.5 Hz), 60.5, 49.6, 43.5, 42.9 (d, *J* = 127.4 Hz), 20.7, 16.2, 16.1, 14.6; HRMS-ESI (M + Na) *m/z* calcd for C₁₇H₂₄NO₆PNa 332.1239, found 332.1239.

(R)-Benzyl-5-(diethoxyphosphoryl)-4-oxopentan-2-ylcarbamate 6b. (Starting from 16 mmol of **5b**) Yellow oil, 3.4 g, yield = 57%; $[\alpha]_D = -26.4$ (c 0.85, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.36–7.22 (m, 5H), 5.26 (brs, 1H), 5.00 (s, 2H), 4.03 (m, 5H), 3.05 (dd, *J* = 23.2, 13.6 Hz, 1H), 2.96 (dd, *J* = 22.7, 13.6 Hz, 1H), 2.85 (dd, *J* = 17.3, 5.8 Hz, 1H), 2.70 (dd, *J* = 17.3, 5.6 Hz, 1H), 1.26 (t, *J* = 6.2 Hz, 6H), 1.16 (d, *J* = 6.7 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 200.7, 155.6, 136.6, 128.5, 128.0, 66.5, 62.7 (d, *J* = 6.6 Hz), 62.6 (d, *J* = 6.8 Hz), 49.5, 43.6, 43.3 (d, *J* = 129.5 Hz), 20.4, 16.3, 16.2; HRMS-ESI (M + Na) *m/z* calcd for C₁₇H₂₆NO₆PNa 394.1395, found 394.1395.

(R and S)-Ethyl-1-(diethoxyphosphoryl)-2-oxoheptan-4-ylcarbamate 6c and 6c'. (Starting from 16 mmol of **5c**) Yellow oil, 3.4 g, yield = 65%; *R* enantiomer: $[\alpha]_D = +43.09$ (c 1.03, CHCl₃), *S* enantiomer: $[\alpha]_D = -42.55$ (c 1.075, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 5.00 (d, *J* = 9.0 Hz, 1H), 4.15–3.97 (m, 6H), 3.90 (m, 1H), 3.09 (dd, *J* = 22.9, 13.5 Hz, 1H), 2.96 (dd, *J* = 22.5, 13.5 Hz, 1H), 2.80 (dd, *J* = 17.2, 5.9 Hz, 1H), 2.74 (dd, *J* = 17.2, 5.3 Hz, 1H), 1.48–1.39 (m, 2H), 1.37–1.29 (m, 2H), 1.27 (dt, *J* = 7.2, 2.0 Hz, 6H), 1.15 (t, *J* = 7.1 Hz, 3H), 0.84 (t, *J* = 7.2 Hz); ¹³C NMR (101 MHz, CDCl₃) δ 200.9, 156.2, 62.7 (d, *J* = 6.5 Hz), 62.6 (d, *J* = 6.6 Hz), 60.6, 48.2, 47.5, 42.9 (d, *J* = 126.6 Hz), 36.7, 19.3, 16.3, 16.2, 14.6, 13.8; HRMS-ESI (M + Na) *m/z* calcd for C₁₄H₂₈NO₆PNa 360.1552, found 360.1562.

(R)-Ethyl-1-(diethoxyphosphoryl)-2-oxododecan-4-ylcarbamate 6d. (Starting from 16 mmol of **5d**) Yellow oil, 4.2 g, yield = 66%; $[\alpha]_D = +30.06$ (c 1.00, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 5.02 (brs, 1H), 4.17–3.99 (m, 6H), 3.95 (m, 1H), 3.07 (dd, *J* = 23.1, 13.5 Hz, 1H), 2.98 (dd, *J* = 22.7, 13.5 Hz, 1H), 2.83 (dd, *J* = 17.4, 5.8 Hz, 1H), 2.75 (dd, *J* = 17.4, 5.2 Hz, 1H), 1.51–1.41 (m, 2H), 1.39–1.23 (m, 12H), 1.21 (t, *J* = 7.2 Hz, 6H), 1.16 (t, *J* = 7.1 Hz, 3H), 0.86 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 200.8, 156.1, 62.9 (d, *J* = 6.6 Hz), 62.8 (d, *J* = 6.7 Hz), 60.7, 48.2, 48.1, 42.7 (d, *J* = 127.1 Hz), 34.4, 31.8, 29.4, 29.3, 29.2, 26.1, 22.6, 16.4, 16.3, 14.6, 14.0; HRMS-ESI (M + Na) *m/z* calcd for C₁₉H₃₈NO₆PNa 430.2334 found 430.2349.

(S)-Ethyl-4-(diethoxyphosphoryl)-3-oxo-1-phenylbutylcarbamate 6e. (Starting from 16 mmol of **5e**) Yellow oil, 3.4 g, yield = 58%; $[\alpha]_D = +1.65$ (c 1.09, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.29–7.21 (m, 4H), 7.16 (m, 1H), 5.72 (s, 1H), 5.09 (dd, *J* = 12.7, 6.7 Hz, 1H), 4.11–3.94 (m, 6H), 3.26 (dd, *J* = 16.9, 7.3 Hz, 1H), 3.04 (dd, *J* = 23.3, 13.1 Hz, 1H), 2.97 (dd, *J* = 16.9, 12.7 Hz, 1H), 2.93 (dd, *J* = 22.9, 13.1

Hz, 1H), 1.22 (td, *J* = 7.1, 1.9 Hz, 6H), 1.13 (t, *J* = 6.7 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 199.9, 155.9, 141.4, 128.6, 127.4, 126.3, 62.8 (d, *J* = 6.2 Hz), 62.6 (d, *J* = 6.5 Hz), 60.8, 51.1, 49.4, 43.3 (d, *J* = 125.5 Hz), 16.2 (d, *J* = 6.1 Hz), 14.6; HRMS-ESI (M + Na) *m/z* calcd for C₁₇H₂₆NO₆PNa 394.1395, found 394.1414.

(S)-tert-Butyl-4-(diethoxyphosphoryl)-3-oxo-1-phenylbutylcarbamate 6f. (Starting from 16 mmol of **5f**) Yellow oil, 3.9 g, yield = 62%; $[\alpha]_D = +3.12$ (c 1.01, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.26–7.12 (m, 4H), 7.16 (m, 1H), 5.44 (brs, 1H), 5.03 (brs, 1H), 4.04–3.95 (m, 4H), 3.20 (dd, *J* = 16.8, 7.4 Hz, 1H), 3.01 (dd, *J* = 23.1, 12.9 Hz, 1H), 2.99 (m, 1H), 2.92 (dd, *J* = 22.7, 12.9 Hz, 1H), 1.32 (s, 9H), 1.21 (td, *J* = 7.1, 1.9 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 200.2, 155.1, 133.6, 130.9, 128.5, 126.3, 66.7, 62.8 (d, *J* = 6.3 Hz), 62.6 (d, *J* = 6.2 Hz), 50.9, 49.6, 41.0 (d, *J* = 127.6 Hz), 16.3 (d, *J* = 5.8 Hz), 16.2 (d, *J* = 5.6 Hz); HRMS-ESI (M + Na) *m/z* calcd for C₁₉H₃₀NO₆PNa 422.1708, found 422.1722.

(S)-Benzyl-4-(diethoxyphosphoryl)-3-oxo-1-phenylbutylcarbamate 6g. (Starting from 16 mmol of **5g**) Yellow oil, 4.5 g, yield = 66%; $[\alpha]_D = +8.66$ (c 1.55, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.31–7.20 (m, 8H), 7.16 (m, 2H), 5.88 (brs, 1H), 5.11 (dd, *J* = 13.1, 7.4 Hz, 1H), 5.04 (d, *J* = 12.3 Hz, 1H), 4.96 (d, *J* = 12.3 Hz, 1H), 4.02–3.89 (m, 4H), 3.27 (dd, *J* = 16.6, 7.4 Hz, 1H), 3.02 (dd, *J* = 23.3, 13.1 Hz, 1H), 2.97 (m, 1H), 2.90 (dd, *J* = 22.6, 13.1 Hz, 1H), 1.21 (t, *J* = 7.0 Hz, 3H), 1.15 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 200.0, 155.6, 141.3, 136.5, 128.6, 128.4, 128.0, 127.5, 126.3, 66.7, 62.9 (d, *J* = 5.3 Hz), 62.6 (d, *J* = 6.4 Hz), 51.3, 49.3, 43.5 (d, *J* = 123.8 Hz), 16.3 (d, *J* = 4.1 Hz), 16.2 (d, *J* = 3.7 Hz); HRMS-ESI (M + Na) *m/z* calcd for C₂₂H₂₈NO₆PNa 456.1552, found 456.1559.

General Procedure for the Synthesis of Enones 7. (R,E)-Ethyl-4-oxo-6-phenylhex-5-en-2-ylcarbamate 7a. To a solution of **6a** (0.5 g, 1.6 mmol, 1 equiv) in THF (7 mL) was added in one time Ba(OH)₂ (0.346 g, 2.0 mmol, 1.25 equiv) at room temperature. After 30 min, a solution of benzaldehyde (0.172 mL, 1.7 mmol, 1.05 equiv) in THF/H₂O 40:1 (7 mL) was slowly added at room temperature. After 1 h, the reaction mixture was quenched with an aqueous solution of ammonium chloride and extracted three times with ethyl acetate. Then the organic layer was dried over Na₂SO₄, concentrated under vacuum, and purified by flash chromatography (eluent, cyclohexane to cyclohexane/EtOAc 8:2) and gave compound **7a** as a white solid (0.40 g, 95%): mp 74 °C; $[\alpha]_D = +9.50$ (c 1.21, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.50 (d, *J* = 16.7 Hz, 1H), 7.47 (dd, *J* = 7.8, 3.0 Hz, 1H), 7.33–7.30 (m, 3H), 6.65 (d, *J* = 16.7 Hz, 1H), 5.14 (s, 1H), 4.14–4.06 (m, 1H), 4.02 (q, *J* = 6.9 Hz, 2H), 2.95 (dd, *J* = 15.9, 4.2 Hz, 1H), 2.71 (dd, *J* = 15.9, 6.5 Hz, 1H), 1.19 (d, *J* = 6.8 Hz, 3H), 1.14 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 198.8, 155.9, 143.4, 134.3, 130.6, 128.9, 128.4, 126.3, 60.6, 46.3, 44.1, 20.5, 14.6; HRMS-ESI (M + Na) calcd for C₁₅H₁₉NO₃Na 284.1263, found 284.1275.

(R,E)-Ethyl-6-(2-nitrophenyl)-4-oxohex-5-en-2-ylcarbamate 7b. $[\alpha]_D = +21.94$ (c 1.015, CHCl₃); yellow solid; starting from 1.6 mmol of **6a**, 0.43 g, yield = 89%; mp 90 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.07 (d, *J* = 8.1 Hz, 1H), 8.01 (d, *J* = 16.1 Hz, 1H), 7.71–7.59 (m, 2H), 7.56 (t, *J* = 7.4 Hz, 1H), 6.58 (d, *J* = 16.1 Hz, 1H), 5.08 (brs, 1H), 4.22–4.08 (m, 3H), 3.04 (dd, *J* = 16.3, 4.7 Hz, 1H), 2.85 (dd, *J* = 16.3, 6.3 Hz, 1H), 1.28 (d, *J* = 6.7 Hz, 3H), 1.22 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 198.3, 155.8, 148.3, 138.7, 133.7, 131.1, 130.8, 130.5, 129.1, 125.1, 60.7, 46.0, 43.9, 20.6, 14.6; HRMS-ESI (M + Na) *m/z* calcd for C₁₅H₁₈N₂O₅Na 329.1113, found 329.1117.

(R,E)-Ethyl-6-(3-nitrophenyl)-4-oxohex-5-en-2-ylcarbamate 7c. $[\alpha]_D = +5.92$ (c 0.995, CHCl₃); yellow solid; starting from 1.6 mmol of **6a**, 0.41 g, yield = 86%; mp 97 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.43 (s, 1H), 8.27 (d, *J* = 8.1 Hz, 1H), 7.87 (d, *J* = 7.7 Hz, 1H), 7.65 (d, *J* = 16.2 Hz, 1H), 7.61 (d, *J* = 7.6 Hz, 1H), 6.85 (d, *J* = 16.2 Hz, 1H), 5.08 (brs, 1H), 4.27–4.04 (m, 3H), 3.07 (dd, *J* = 16.1, 3.3 Hz, 1H), 2.84 (dd, *J* = 16.1, 6.6 Hz, 1H), 1.31 (d, *J* = 6.7 Hz, 3H), 1.25 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 198.3, 155.6, 148.5, 140.2, 136.0, 133.9, 130.0, 128.6, 124.8, 122.6, 60.8, 47.0, 44.0, 20.5, 14.6; HRMS-ESI (M + Na) *m/z* calcd for C₁₅H₁₈N₂O₅Na 329.1113, found 329.1125.

(*R,E*)-Ethyl-6-(4-nitrophenyl)-4-oxohex-5-en-2-ylcarbamate **7d**. $[\alpha]_D^{25} = +16.42$ (c 0.52, CHCl₃); yellow solid; starting from 1.6 mmol of **6a**, 0.44 g, yield = 91%; mp 98 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.19 (d, *J* = 8.8 Hz, 2H), 7.62 (d, *J* = 8.5 Hz, 2H), 7.53 (d, *J* = 16.2 Hz, 1H), 6.75 (d, *J* = 16.2 Hz, 1H), 4.97 (s, 1H), 4.10 (m, 1H), 4.03 (q, *J* = 7.0 Hz, 2H), 2.99 (dd, *J* = 15.6, 3.5 Hz, 1H), 2.74 (dd, *J* = 15.6, 6.5 Hz, 1H), 1.21 (d, *J* = 6.8 Hz, 1H), 1.15 (t, *J* = 7.0 Hz, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 198.0, 155.9, 140.5, 140.1, 128.5, 128.9, 124.8, 124.2, 60.8, 47.1, 44.0, 20.5, 14.6; HRMS-ESI (M + Na) *m/z* calcd for C₁₅H₁₈N₂O₅Na 329.1113, found 329.1119.

(*R,E*)-Ethyl-6-(4-methoxyphenyl)-4-oxohex-5-en-2-ylcarbamate **7e**. $[\alpha]_D^{25} = +6.1$ (c 1.055, CHCl₃); yellow solid; starting from 1.6 mmol of **6a**, 0.37 g, yield = 79%; mp 108 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.54 (d, *J* = 16.1 Hz, 1H), 7.50 (d, *J* = 7.9 Hz, 2H), 6.91 (d, *J* = 7.9 Hz, 1H), 6.60 (d, *J* = 16.1 Hz, 1H), 5.21 (brs, 1H), 4.22–4.07 (m, 3H), 3.83 (s, 3H), 2.99 (dd, *J* = 15.8, 4.4 Hz, 1H), 2.76 (dd, *J* = 15.8, 5.8 Hz, 1H), 1.25 (d, *J* = 6.8 Hz, 1H), 1.21 (t, *J* = 7.0 Hz, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 198.8, 161.9, 155.9, 143.2, 130.3, 127.1, 124.3, 114.6, 60.8, 55.5, 46.2, 44.4, 20.6, 14.7; HRMS-ESI (M + Na) *m/z* calcd for C₁₆H₂₁NO₄Na 314.1368, found 314.1371.

(*R,E*)-Ethyl-6-(2-bromophenyl)-4-oxohex-5-en-2-ylcarbamate **7f**. $[\alpha]_D^{25} = +18.2$ (c 1.175, CHCl₃); yellow solid; starting from 1.6 mmol of **6a**, 0.53 g, yield = 91%; mp 65 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.86 (d, *J* = 16.2 Hz, 1H), 7.56 (d, *J* = 7.8 Hz, 1H), 7.56 (d, *J* = 7.8 Hz, 1H), 7.27 (t, *J* = 7.8 Hz, 1H), 7.17 (td, *J* = 7.8, 1.5 Hz, 1H), 6.57 (d, *J* = 16.2 Hz, 1H), 5.08 (brs, 1H), 4.15–3.88 (m, 3H), 2.96 (dd, *J* = 16.4, 4.8 Hz, 1H), 2.79 (dd, *J* = 16.3, 6.2 Hz, 1H), 1.22 (d, *J* = 6.8 Hz, 3H), 1.16 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 198.8, 158.1, 145.0, 141.6, 133.5, 131.5, 129.0, 127.8, 60.7, 46.5, 44.0, 20.6, 14.6; HRMS-ESI (M + Na) *m/z* calcd for C₁₅H₁₈BrNO₃Na 362.0368, found 362.0371.

(*R,E*)-Ethyl-6-(4-bromophenyl)-4-oxo-hex-5-en-2-ylcarbamate **7g**. $[\alpha]_D^{25} = +4.0$ (c 1.03, CHCl₃); yellow solid; starting from 1.6 mmol of **6a**, 0.48 g, yield = 89%; mp 90 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.53 (d, *J* = 8.6 Hz, 2H), 7.50 (d, *J* = 16.2 Hz, 1H), 7.41 (d, *J* = 8.5 Hz, 1H), 6.70 (d, *J* = 16.2 Hz, 1H), 5.10 (s, 1H, NH), 4.14 (m, 1H), 4.09 (q, *J* = 7.1 Hz, 1H), 3.01 (dd, *J* = 15.5, 3.4 Hz, 1H), 2.77 (dd, *J* = 15.5, 6.6 Hz, 1H), 1.27 (d, *J* = 6.8 Hz, 1H), 1.23 (t, *J* = 7.1, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 198.4, 156.0, 141.9, 133.2, 132.2, 129.7, 126.7, 124.9, 60.7, 50.0, 46.5, 44.1, 20.5, 14.6; HRMS-ESI (M + Na) *m/z* calcd for C₁₅H₁₈BrNO₃Na 362.0368, found 362.0380.

(*R,E*)-Ethyl-6-(2-chloro-5-nitrophenyl)-4-oxo-hex-5-en-2-ylcarbamate **7h**. $[\alpha]_D^{25} = +15.06$ (c 1.06, CHCl₃); yellow solid; starting from 1.6 mmol of **6a**, 0.52 g, yield = 95%; mp 149 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.05 (d, *J* = 8.6 Hz, 1H), 7.97 (d, *J* = 16.0 Hz, 1H), 7.62 (s, 1H), 7.51 (d, *J* = 8.6, 1H), 6.57 (d, *J* = 16.0 Hz, 1H), 5.09 (brs, 1H), 4.23–4.03 (m, 3H), 3.05 (dd, *J* = 16.5, 5.2 Hz, 1H), 2.84 (dd, *J* = 16.5, 6.5 Hz, 1H), 1.28 (d, *J* = 6.9 Hz, 3H), 1.21 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 198.0, 158.9, 146.5, 140.4, 137.7, 132.9, 131.9, 130.5, 129.3, 126.7, 60.9, 46.5, 44.1, 20.7, 14.7; HRMS-ESI (M + Na) *m/z* calcd for C₁₅H₁₇ClN₂O₅Na 363.0724, found 363.0717.

(*R,E*)-Ethyl-4-oxo-6-(pyridin-3-yl)hex-5-en-2-ylcarbamate **7i**. $[\alpha]_D^{25} = +10.25$ (c 0.865, CHCl₃); white solid; starting from 1.6 mmol of **6a**, 0.36 g, yield = 87%; mp 90 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.71 (d, *J* = 1.6 Hz, 1H), 8.56 (d, *J* = 4.7 Hz, 1H), 7.82 (dt, *J* = 7.9, 1.6, 1H), 7.51 (d, *J* = 16.3 Hz, 1H), 7.30 (dd, *J* = 7.9, 4.9 Hz, 1H), 6.72 (d, *J* = 16.3 Hz, 1H), 5.03 (brs, 1H), 4.20–3.70 (m, 3H), 2.98 (dd, *J* = 16.1, 4.0 Hz, 1H), 2.74 (dd, *J* = 16.1, 6.7 Hz, 1H), 1.21 (d, *J* = 6.9 Hz, 3H), 1.16 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 198.1, 155.9, 151.1, 149.9, 139.4, 134.5, 130.2, 128.0, 123.8, 60.7, 46.7, 44.1, 20.5, 14.6; HRMS-ESI (M + Na) *m/z* calcd for C₁₄H₁₈N₂O₃Na 285.1215, found 285.1210.

(*R,2E,4E*)-Ethyl-8-(ethoxycarbonylamino)-6-oxo-nona-2,4-dienoate **7j**. $[\alpha]_D^{25} = +17.6$ (c 0.695, CHCl₃); viscous yellow oil; starting from 1.6 mmol of **6a**, 0.38 g, yield = 85%; ¹H NMR (400 MHz, CDCl₃) δ 7.24 (dd, *J* = 14.9, 11.4 Hz, 1H), 7.14 (dd, *J* = 15.1, 11.4 Hz, 1H), 6.35 (d, *J* = 15.0 Hz, 1H), 6.19 (d, *J* = 15.0 Hz, 1H), 4.97 (s, 1H), 4.17 (q, *J* = 7.1 Hz, 2H), 4.05–4.00 (m, 3H), 2.89 (d, *J* = 12.5 Hz, 1H), 2.67 (dd, *J* = 16.3, 6.5 Hz, 1H), 1.25 (t, *J* = 7.1 Hz, 3H), 1.18 (d, *J* = 6.8 Hz, 3H), 1.16 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz,

CDCl₃) δ 198.4, 165.7, 155.8, 141.1, 139.1, 135.3, 129.5, 60.9, 60.7, 46.7, 43.9, 20.4, 14.6, 14.2; HRMS-ESI (M + Na) *m/z* calcd for C₁₄H₂₁NO₅Na 306.1317, found 306.1331.

(*R*)-Ethyl-6-(ethoxycarbonylamino)-4-oxohept-2-enoate **7k**. Mixture of *Z* and *E* isomers (*Z/E* = 60:40); colorless oil; starting from 1.6 mmol of **6a**, 0.22 g, yield = 53%; ¹H NMR (400 MHz, CDCl₃) δ 6.96 (d, *J* = 16.0 Hz, 1H), 6.62 (d, *J* = 16.0 Hz, 1H), 6.43 (d, *J* = 12.0 Hz, 1H), 5.97 (d, *J* = 12.0 Hz, 1H), 5.18 (s, 1H), 5.12 (s, 1H), 4.25–4.11 (m, 6H), 2.91 (d, *J* = 15.0 Hz, 1H), 2.81 (dd, *J* = 16.9, 5.7 Hz, 1H), 2.78–2.67 (m, 1H), 1.27–1.13 (m, 12H); ¹³C NMR (101 MHz, CDCl₃) (mixture of *Z* and *E*) δ 201.1, 197.3, 164.4, 164.2, 155.7, 155.0, 140.6, 138.3, 130.4, 124.0, 60.5, 60.3, 59.8, 59.7, 47.3, 46.0, 42.7, 42.4, 19.6, 19.4, 13.6, 13.1, 13.0; HRMS-ESI (M + Na) *m/z* calcd for C₁₂H₁₉NO₅Na 280.1161, found 280.1163.

(*R,E*)-Ethyl-4-oxo-non-5-en-2-ylcarbamate **7l**. $[\alpha]_D^{25} = +12.13$ (c 1.025, CHCl₃); colorless oil; starting from 1.6 mmol of **6a**, 0.34 g, yield = 95%; ¹H NMR (400 MHz, CDCl₃) δ 6.84 (dt, *J* = 15.9, 7.2 Hz, 1H), 6.07 (dd, *J* = 15.9, 1.5 Hz, 1H), 5.13 (brs, 1H), 4.11–4.02 (m, 3H), 2.87 (dd, *J* = 16.1, 4.4 Hz, 1H), 2.65 (dd, *J* = 16.1, 6.4 Hz, 1H), 2.19 (td, *J* = 7.2, 1.5 Hz, 2H), 1.49 (qd, *J* = 7.2 Hz, 2H), 1.22 (t, *J* = 7.2 Hz, 3H), 1.21 (d, *J* = 6.8 Hz, 3H), 0.92 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 199.1, 156.0, 148.5, 130.9, 60.7, 45.5, 44.2, 34.6, 21.4, 20.6, 14.7, 13.8; HRMS-ESI (M + Na) *m/z* calcd for C₁₂H₂₁NO₃Na 250.1419, found 250.1421.

(*R,E*)-Ethyl-4-oxo-tetradec-5-en-2-ylcarbamate **7m**. $[\alpha]_D^{25} = +10.71$ (c 1.025, CHCl₃); yellow oil; starting from 1.6 mmol of **6a**, 0.42 g, yield = 88%; ¹H NMR (400 MHz, CDCl₃) δ 6.79 (dt, *J* = 15.8, 6.9 Hz, 1H), 6.01 (d, *J* = 15.8 Hz, 1H), 5.16 (s, 1H), 4.07–4.03 (m, 1H, 3H), 2.86 (dd, *J* = 16.0, 4.0 Hz, 1H), 2.63 (dd, *J* = 16.0, 6.4 Hz, 1H), 2.14 (q, *J* = 6.9 Hz, 2H), 1.45–1.32 (m, 2H), 1.30–1.18 (m, 15H), 1.15 (d, *J* = 7.1 Hz, 3H), 0.81 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 199.1, 155.9, 148.7, 130.7, 60.7, 45.5, 44.1, 32.6, 31.9, 29.5, 29.4, 29.3, 29.2, 28.1, 22.7, 20.5, 14.7, 14.2; HRMS-ESI (M + Na) *m/z* calcd for C₁₇H₃₁NO₃Na 320.2202, found 320.2209.

(*R,E*)-Benzyl-4-oxo-6-phenyl-hex-5-en-2-ylcarbamate **7n**. $[\alpha]_D^{25} = +2.56$ (c 1.95, CHCl₃); white solid; starting from 1.3 mmol of **6b**, 0.40 g, yield = 96%; mp 96 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.50 (d, *J* = 16.1 Hz, 1H), 7.48–7.46 (m, 2H), 7.33–7.19 (m, 8H), 6.64 (d, *J* = 16.1 Hz, 1H), 5.23 (s, 1H), 5.10–4.96 (m, 2H), 4.18–4.06 (m, 1H), 2.97 (d, *J* = 15.7 Hz, 1H), 2.73 (dd, *J* = 15.7, 5.6 Hz, 1H), 1.21 (d, *J* = 6.2 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 199.5, 155.6, 143.1, 134.0, 130.7, 129.0, 128.5, 128.4, 128.1, 126.3, 66.6, 46.1, 44.3, 20.5; HRMS-ESI (M + Na) *m/z* calcd for C₂₀H₂₁NO₃Na 346.1419, found 346.1424.

(*R,E*)-Benzyl-4-oxo-pentadec-5-en-2-ylcarbamate **7o**. $[\alpha]_D^{25} = -9.86$ (c 0.975, CHCl₃); yellow oil; starting from 1.3 mmol of **6b**, 0.38 g, yield = 78%; ¹H NMR (400 MHz, CDCl₃) δ 7.42–7.30 (m, 5H), 6.87 (dt, *J* = 16.0, 6.7 Hz, 1H), 6.09 (d, *J* = 16.0 Hz, 1H), 5.29 (brs, 1H), 5.10 (brs, 2H), 4.13 (m, 1H), 2.91 (dd, *J* = 16.1, 3.1 Hz, 1H), 2.69 (dd, *J* = 16.1, 6.1 Hz, 1H), 2.22 (dt, *J* = 6.7, 7.1 Hz, 2H), 1.52–1.41 (m, 2H), 1.36–1.27 (m, 12H), 1.25 (d, *J* = 6.8 Hz, 3H), 0.90 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 200.4, 155.6, 148.7, 139.5, 130.5, 128.5, 128.1, 128.0, 66.5, 45.2, 44.2, 32.5, 31.8, 29.5, 29.4, 29.3, 29.2, 28.04, 22.6, 20.4, 14.1; HRMS-ESI (M + H)⁺ *m/z* calcd for C₂₃H₃₆NO₃ 374.2695, found 374.2706.

(*R,E*)-Benzyl-4-oxo-heptadec-5-en-2-ylcarbamate **7p**. $[\alpha]_D^{25} = -9.80$ (c 1.015, CHCl₃); yellow oil; starting from 1.3 mmol of **6b**, 0.45 g, yield = 91%; ¹H NMR (400 MHz, CDCl₃) δ 7.42–7.30 (m, 5H), 6.87 (dt, *J* = 15.9, 7.2 Hz, 1H), 6.06 (d, *J* = 15.9 Hz, 1H), 5.31 (brs, 1H), 5.08 (brs, 2H), 4.10 (m, 1H), 2.89 (d, *J* = 15.6 Hz, 1H), 2.66 (dd, *J* = 15.6, 5.3 Hz, 1H), 2.19 (q, *J* = 7.2 Hz, 2H), 1.47–1.40 (m, 6H), 1.33–1.21 (m, 15H), 0.88 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 199.1, 155.7, 148.9, 136.7, 130.6, 128.6, 128.1, 66.6, 45.3, 44.3, 32.6, 32.0, 29.7, 29.6, 29.5, 29.4, 29.3, 28.1, 27.0, 22.8, 20.5, 14.2; HRMS-ESI (M + H)⁺ *m/z* calcd for C₂₅H₄₀NO₃ 402.3008, found 402.3015.

(*E*)-Ethyl-6-oxo-8-phenyl-oct-7-en-4-ylcarbamate **7q** and **7q'**. *R* enantiomer: $[\alpha]_D^{25} = +21.87$ (c 0.97, CHCl₃); *S* enantiomer: $[\alpha]_D^{25} = -21.32$ (c 0.76 CHCl₃); white solid; starting from 1.5 mmol of **6c**, 0.39 g, yield = 91%; mp 96 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.48

(d, $J = 16.8$ Hz, 1H), 7.45–7.3 (m, 2H), 7.33–7.30 (m, 3H), 6.64 (d, $J = 16.8$ Hz, 1H), 5.06 (brs, 1H), 4.09–3.91 (m, 3H), 2.90 (dd, $J = 17.2$, 6.0 Hz, 1H), 2.78 (dd, $J = 17.2$, 5.5 Hz, 1H), 1.62–1.29 (m, 4H), 1.17 (t, $J = 6.9$ Hz, 3H), 0.85 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 199.0, 156.2, 143.2, 134.3, 130.6, 128.9, 128.4, 126.3, 60.6, 48.1, 44.8, 36.5, 19.5, 14.6, 13.8; HRMS-ESI ($M + \text{Na}$) m/z calcd for $\text{C}_{17}\text{H}_{23}\text{NO}_3\text{Na}$ 312.1576, found 312.1585.

(*R,E*)-Ethyl-8-(4-nitrophenyl)-6-oxo-oct-7-en-4-ylcarbamate **7r**. $[\alpha]_{\text{D}} = +13.8$ (c 0.985, CHCl_3); yellow solid; starting from 1.5 mmol of **6c**, 0.42 g, yield = 84%; mp 102 °C; ^1H NMR (400 MHz, CDCl_3) δ 8.25 (d, $J = 8.1$ Hz, 2H), 7.70 (d, $J = 8.1$ Hz, 2H), 7.60 (d, $J = 16.2$ Hz, 1H), 6.83 (d, $J = 16.2$ Hz, 1H), 5.01 (s, 1H), 4.14–4.01 (m, 3H), 3.01 (d, $J = 16.2$ Hz, 1H), 2.85 (dd, $J = 16.2$, 5.7 Hz, 1H), 1.64–1.49 (m, 2H), 1.47–1.30 (m, 2H), 1.22 (t, $J = 6.8$ Hz, 3H), 0.92 (t, $J = 7.2$ Hz, 1H); ^{13}C NMR (101 MHz, CDCl_3) δ 198.5, 156.2, 140.6, 140.0, 129.5, 128.9, 124.2, 124.2, 60.8, 48.1, 45.8, 36.6, 19.5, 14.6, 13.8; HRMS-ESI ($M + \text{Na}$) m/z calcd for $\text{C}_{17}\text{H}_{22}\text{N}_2\text{O}_5\text{Na}$ 357.1426, found 357.1420.

(*R,E*)-Ethyl-3-oxo-1-phenyl-tridec-1-en-5-ylcarbamate **7s**. $[\alpha]_{\text{D}} = +17.26$ (c 1.015, CHCl_3); white solid; starting from 1.2 mmol of **6d**, 0.40 g, yield = 94%; mp 76 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.58 (d, $J = 16.1$ Hz, 1H), 7.57–7.52 (m, 2H), 7.44–7.36 (m, 3H), 6.74 (d, $J = 16.1$ Hz, 1H), 5.23 (d, $J = 7.4$ Hz, 1H), 4.21–3.94 (m, 3H), 3.01 (d, $J = 15.2$ Hz, 1H), 2.84 (dd, $J = 15.2$, 3.8 Hz, 1H), 1.69–1.49 (m, 2H), 1.48–1.11 (m, 15H), 0.85 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 199.0, 156.2, 143.2, 134.3, 130.6, 129.6, 128.9, 128.4, 126.3, 60.6, 48.4, 44.9, 34.4, 31.8, 29.5, 29.2, 26.3, 22.6, 14.6, 14.1; HRMS-ESI ($M + \text{Na}$) m/z calcd for $\text{C}_{22}\text{H}_{33}\text{NO}_3\text{Na}$ 382.2358, found 382.2364.

(*R,E*)-Ethyl-4-oxo-tetradec-2-en-6-ylcarbamate **7t**. $[\alpha]_{\text{D}} = +17.44$ (c 1.12, CHCl_3); white solid; starting from 1.2 mmol of **6d**, 0.27 g, yield = 78%; mp 59 °C; ^1H NMR (400 MHz, CDCl_3) δ 6.87 (dt, $J = 15.8$, 6.8, 1H), 6.10 (d, $J = 15.8$, 1.6 Hz, 1H), 5.1 (brs, 1H), 4.07 (q, $J = 7.2$ Hz, 2H), 3.91 (m, 1H), 2.84 (dd, $J = 15.8$, 4.2 Hz, 1H), 2.67 (dd, $J = 15.8$, 5.5 Hz, 1H), 1.90 (d, 3H, $J = 6.8$ Hz), 1.55–1.46 (m, 2H), 1.33–1.20 (m, 15H), 0.86 (t, $J = 6.9$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 199.2, 156.3, 143.6, 132.4, 60.7, 48.5, 44.0, 34.5, 31.9, 29.6, 29.5, 29.4, 26.4, 22.8, 18.5, 14.7, 14.2; HRMS-ESI ($M + \text{Na}$) m/z calcd for $\text{C}_{17}\text{H}_{31}\text{NO}_3\text{Na}$ 320.2202, found 320.2207.

(*R,E*)-Ethyl-6-oxo-hexadec-4-en-8-ylcarbamate **7u**. $[\alpha]_{\text{D}} = +12.1$ (c 1.05, CHCl_3); colorless oil; starting from 1.2 mmol of **6d**, 0.37 g, yield = 95%; ^1H NMR (400 MHz, CDCl_3) δ 6.83 (dt, $J = 15.9$, 7.2 Hz, 1H), 6.07 (dd, $J = 15.9$, 1.4 Hz, 1H), 5.11 (brs, 1H), 4.07 (qd, $J = 6.9$, 2H), 3.91 (m, 1H), 2.85 (dd, $J = 16.3$, 4.5 Hz, 1H), 2.69 (dd, $J = 16.3$, 5.7 Hz, 1H), 2.18 (dd, $J = 7.2$, 1.4 Hz, 1H), 1.53–1.44 (m, 2H), 1.49 (qd, $J = 7.2$ Hz, 2H), 1.29–1.19 (m, 15H), 0.92 (t, $J = 7.2$ Hz, 3H), 0.86 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 199.4, 156.3, 148.3, 130.9, 60.7, 46.5, 44.1, 34.6, 34.5, 31.9, 29.6, 29.5, 29.4, 26.4, 22.8, 21.4, 14.7, 14.2, 13.8; HRMS-ESI ($M + \text{Na}$) m/z calcd for $\text{C}_{19}\text{H}_{35}\text{NO}_3\text{Na}$ 348.2515, found 348.2525.

(*S,E*)-Ethyl-3-oxo-1,5-diphenyl-pent-4-enylcarbamate **7v**. $[\alpha]_{\text{D}} = +6.6$ (c 0.94, CHCl_3); yellow oil; starting from 1.3 mmol of **6e**, 0.35 g, yield = 84%; ^1H NMR (400 MHz, CDCl_3) δ 7.43 (d, $J = 16.2$ Hz, 1H), 7.41–7.11 (m, 10H), 6.59 (d, $J = 16.2$ Hz, 1H), 5.74 (brs, 1H), 5.16 (m, 1H), 4.01 (q, $J = 6.3$ Hz, 1H), 3.26 (dd, $J = 15.9$ Hz, 1H), 3.06 (dd, $J = 15.9$, 5.0 Hz, 1H), 1.13 (t, $J = 6.3$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 197.9, 155.9, 143.6, 134.2, 130.7, 128.9, 128.6, 128.4, 127.5, 126.3, 126.0, 61.0, 51.7, 46.1, 14.6; HRMS-ESI ($M + \text{Na}$) m/z calcd for $\text{C}_{20}\text{H}_{21}\text{NO}_3\text{Na}$ 346.1419, found 346.1414.

(*S,E*)-Ethyl-3-oxo-1-phenyl-hex-4-enylcarbamate **7w**. $[\alpha]_{\text{D}} = -13.3$ (c 0.715, CHCl_3); yellow oil; starting from 1.3 mmol of **6e**, 0.27 g, yield = 80%; ^1H NMR (400 MHz, CDCl_3) δ 7.32–7.12 (m, 5H), 6.75 (dq, $J = 15.9$, 6.8 Hz, 1H), 5.96 (dd, $J = 15.9$, 1.5 Hz, 1H), 5.68 (brs, 1H), 5.09 (m, 1H), 4.01 (q, $J = 6.9$ Hz, 2H), 3.12 (dd, $J = 16.6$, 5.8 Hz, 1H), 2.93 (dd, $J = 16.6$, 6.8 Hz, 1H), 1.79 (dd, $J = 6.8$, 1.5 Hz, 3H), 1.10 (t, $J = 6.9$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 200.2, 158.2, 146.2, 143.7, 134.2, 130.8, 129.6, 128.5, 63.2, 53.9, 45.5, 20.6, 16.8; HRMS-ESI ($M + \text{Na}$) m/z calcd for $\text{C}_{15}\text{H}_{19}\text{NO}_3\text{Na}$ 284.1263, found 284.1275.

(*S,E*)-Ethyl-3-oxo-1-phenyl-oct-4-enylcarbamate **7x**. $[\alpha]_{\text{D}} = -9.11$ (c 1.14, CHCl_3); colorless oil; starting from 1.3 mmol of **6e**, 0.36 g, yield = 95%; ^1H NMR (400 MHz, CDCl_3) δ 7.35–7.21 (m, 5H), 6.80 (dt, $J = 15.9$, 6.9 Hz, 1H), 6.05 (td, $J = 15.9$, 1.5, 1H), 5.75 (s, 1H), 5.15 (dd, $J = 5.8$, 3.1 Hz, 1H), 4.11 (q, $J = 7.1$ Hz, 2H), 3.23 (dd, $J = 16.1$, 3.1 Hz, 1H), 3.04 (dd, $J = 16.1$, 5.8 Hz, 1H), 2.16 (qd, $J = 6.9$, 1.5 Hz, 2H), 1.47 (sex, $J = 6.8$ Hz, 2H), 1.23 (t, $J = 7.1$ Hz, 3H), 0.92 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 198.3, 156.1, 148.8, 141.6, 130.6, 128.6, 127.4, 126.4, 61.0, 51.8, 45.3, 34.6, 21.3, 14.6, 13.7; HRMS-ESI ($M + \text{Na}$) m/z calcd for $\text{C}_{17}\text{H}_{23}\text{NO}_3\text{Na}$ 312.1576, found 312.1576.

(*S,E*)-Ethyl-3-oxo-1-phenyl-tridec-4-enylcarbamate **7y**. $[\alpha]_{\text{D}} = -1.22$ (c 0.995, CHCl_3); white solid; starting from 1.3 mmol of **6e**, 0.41 g, yield = 89%; mp 60 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.23–7.17 (m, 4H), 7.13 (m, 1H), 6.70 (dt, $J = 15.9$, 6.9 Hz, 1H), 5.93 (d, $J = 15.9$ Hz, 1H), 5.88 (brs, 1H), 5.07 (m, 1H), 3.98 (q, $J = 7.1$ Hz, 2H), 3.09 (d, $J = 16.1$ Hz, 1H), 2.89 (dd, $J = 16.1$, 5.7 Hz, 1H), 2.06 (q, $J = 6.9$ Hz, 2H), 1.39–1.29 (m, 2H), 1.27–1.13 (m, 10H), 1.10 (t, $J = 7.1$ Hz, 3H), 0.79 (t, $J = 6.9$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 198.2, 156.0, 148.9, 141.7, 130.3, 128.5, 127.3, 126.3, 60.8, 51.6, 45.3, 32.5, 31.8, 29.4, 29.3, 29.2, 29.1, 27.9, 22.6, 14.5, 14.1; HRMS-ESI ($M + \text{Na}$) m/z calcd for $\text{C}_{22}\text{H}_{33}\text{NO}_3\text{Na}$ 382.2358, found 382.2362.

(*S,E*)-tert-Butyl-3-oxo-1-phenyl-hex-4-enylcarbamate **7z**. $[\alpha]_{\text{D}} = -10.68$ (c 1.015, CHCl_3); white solid; starting from 1.2 mmol of **6f**, 0.28 g, yield = 81%; mp 94 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.32–7.18 (m, 5H), 6.84 (dq, $J = 15.9$, 6.8 Hz, 1H), 6.06 (d, $J = 15.9$, 1.6 Hz, 1H), 5.55 (brs, 1H), 5.09 (m, 1H), 3.12 (d, $J = 16.6$ Hz, 1H), 2.98 (dd, $J = 16.6$, 5.6 Hz, 1H), 1.85 (dd, $J = 6.8$, 1.6 Hz, 3H), 1.40 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 198.2, 155.3, 144.0, 132.1, 128.7, 127.4, 126.4, 51.5, 45.6, 28.5, 18.5; HRMS-ESI ($M + \text{Na}$) m/z calcd for $\text{C}_{17}\text{H}_{23}\text{NO}_3\text{Na}$ 312.1576, found 312.1591.

(*S,E*)-Benzyl-3-oxo-1-phenyl-hex-4-enylcarbamate **1**. $[\alpha]_{\text{D}} = -5.34$ (c 1.05, CHCl_3); white solid; starting from 1.1 mmol of **6g**, 0.3 g, yield = 96%; mp 60 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.42–7.17 (m, 5H), 6.80 (dq, $J = 15.8$, 6.8 Hz, 1H), 6.05 (d, $J = 15.8$ Hz, 1H), 5.88 (brs, 1H), 5.18 (dd, $J = 6.2$, 5.6 Hz, 1H), 5.10 (d, $J = 12.3$ Hz, 1H), 5.05 (d, $J = 12.3$ Hz, 1H), 3.19 (dd, $J = 16.2$, 5.6 Hz, 1H), 3.00 (dd, $J = 16.2$, 6.2 Hz, 1H), 1.85 (d, $J = 6.8$ Hz, 1H); ^{13}C NMR (101 MHz, CDCl_3) δ 197.8, 155.7, 144.0, 136.4, 136.2, 131.9, 128.7, 128.6, 128.5, 128.0, 126.4, 126.3, 66.8, 51.8, 45.1, 18.3; HRMS-ESI ($M + \text{Na}$) m/z calcd for $\text{C}_{21}\text{H}_{21}\text{NO}_3\text{Na}$ 346.1419, found 346.1426.

General Procedure for the Synthesis of Piperidines 8l–s/9l–s. (*2S,6R*)-Ethyl-4,4-dimethoxy-6-methyl-2-[(4-nitrophenyl)]-piperidine-1-carboxylate **8m** (*trans isomer*) and (*2R,6R*)-Ethyl-4,4-dimethoxy-6-methyl-2-[(4-nitrophenyl)]piperidine-1-carboxylate **9m** (*cis isomer*). In a one neck flask, to compound **7d** (0.1 g, 0.32 mmol 1 equiv) were added successively, trimethyl orthoformate (1.75 mL, 1.60 mmol, 5 equiv) and *p*-toluenesulphonic acid (5.5 mg, 0.32 mmol, 1 equiv). The reaction was followed by TLC, and after 0.5 h ethyl acetate was added to the crude mixture, followed by a saturated solution of NaHCO_3 and extraction twice with ethyl acetate. The organic layer was dried and concentrated under vacuum before being purified by flash chromatography (eluent, cyclohexane to cyclohexane/ EtOAc 8:2) to yield a mixture of both isomer **8m** and **9m** (86 mg, 81% yield) in a ratio of **8m/9m** 89/11 in favor of the *trans isomer* **8m**.

8m. ^1H NMR (400 MHz, C_6D_6) δ 7.87 (d, $J = 8.8$ Hz, 2H), 6.84 (d, $J = 8.8$ Hz, 2H), 4.98 (t, $J = 5.3$ Hz, 1H), 4.26 (m, 1H), 4.09–3.92 (m, 2H), 2.87 (s, 3H), 2.58 (s, 3H), 2.03 (d, $J = 5.3$ Hz, 2H), 1.67 (dd, $J = 3.5$, 14.4 Hz, 1H), 1.61 (dd, $J = 5.5$, 14.4 Hz, 1H), 1.37 (d, $J = 6.7$ Hz, 3H), 0.86 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, C_6D_6) δ 155.7, 150.8, 146.8, 123.3, 126.6, 98.2, 61.2, 53.7, 47.6, 47.2, 47.0, 37.6, 36.7, 20.6, 14.4; HRMS ($M + \text{H}$)⁺ ion by direct probe calcd for $\text{C}_{17}\text{H}_{25}\text{N}_2\text{O}_6$ 353.1713, found 353.1699.

9m. ^1H NMR (400 MHz, C_6D_6) δ 7.84 (d, $J = 8.8$ Hz, 2H), 7.06 (d, $J = 8.8$ Hz, 2H), 5.19 (dd, $J = 6.6$, 6.9 Hz, 1H), 4.43 (m, 1H), 4.09–3.92 (m, 2H), 2.98 (s, 3H), 2.79 (s, 3H), 2.01 (dd, $J = 6.9$, 14.1 Hz, 1H), 1.94 (dd, $J = 6.6$, 14.1 Hz, 1H), 1.79 (dd, $J = 6.7$, 14.2 Hz, 1H), 1.43 (dd, $J = 5.9$, 14.2 Hz, 1H), 1.16 (d, $J = 6.7$ Hz, 3H), 0.87 (t, $J =$

7.1 Hz, 3H); ^{13}C NMR (101 MHz, C_6D_6) δ 155.7, 151.9, 146.8, 123.8, 126.9, 98.6, 61.5, 54.0, 48.2, 47.2, 47.0, 36.9, 34.0, 23.0, 14.4.

(6*R*,2*S*)-Ethyl-4,4-dimethoxy-6-methyl-2-(pyridin-3-yl)piperidine-1-carboxylate **8k** and (6*R*,2*R*)-Ethyl-4,4-dimethoxy-6-methyl-2-(pyridin-3-yl)piperidine-1-carboxylate **9k**. Yellow oil, starting from 0.8 mmol of **7i**, 0.20 g, yield = 81% in a ratio of **8k**/**9k** 85/15 in favor of the *trans* isomer **8k**.

8k. ^1H NMR (400 MHz, CDCl_3) δ 8.40 (m, 1H), 8.38 (d, J = 4.5 Hz, 1H), 7.44 (d, J = 7.3 Hz, 1H), 7.16 (dd, J = 7.3, 4.5 Hz, 1H), 5.09 (t, J = 5.3 Hz, 1H), 4.26 (m, 1H), 4.06–3.94 (m, 2H), 3.11 (s, 3H), 2.85 (s, 3H), 2.35 (dd, J = 14.4, 5.3 Hz, 1H), 2.29 (dd, J = 14.4, 5.3 Hz, 1H), 1.88 (dd, J = 14.4, 5.3 Hz, 1H), 1.82 (dd, J = 14.4, 5.5 Hz, 1H), 1.32 (d, J = 6.8 Hz, 3H), 1.04 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 156.1, 147.7, 147.5, 137.9, 133.5, 122.9, 98.2, 61.3, 51.7, 47.8, 47.6, 46.8, 37.3, 36.6, 20.6, 14.4.

9k. ^1H NMR (400 MHz, CDCl_3) δ 8.53 (m, 1H), 8.37 (d, J = 4.5 Hz, 1H), 7.55 (d, J = 7.3 Hz, 1H), 7.18 (dd, J = 7.3, 4.5 Hz, 1H), 5.16 (dd, J = 7.1, 6.4 Hz, 1H), 4.35 (m, 1H), 4.06–3.94 (m, 2H), 3.16 (s, 3H), 3.05 (s, 3H), 2.27 (m, 1H), 2.15 (dd, J = 14.6, 6.4 Hz, 1H), 2.00 (dd, J = 14.4, 6.9 Hz, 1H), 1.66 (dd, J = 14.4, 5.5 Hz, 1H), 1.22 (d, J = 6.9 Hz, 3H), 1.04 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 155.9, 147.4, 147.9, 138.1, 133.8, 123.1, 98.4, 61.5, 52.3, 47.9, 47.6, 47.3, 37.4, 36.7, 22.9, 14.4; HRMS-ESI ($\text{M} + \text{Na}$) m/z calcd for $\text{C}_{16}\text{H}_{24}\text{N}_2\text{O}_4\text{Na}$ 331.1634, found 331.1634.

(2*S*,6*R*)-ethyl-4,4-dimethoxy-6-propyl-2-[(4-nitrophenyl)]piperidine-1-carboxylate **8l** and **9l**. Yellow oil, starting from 0.6 mmol of **7r**, 0.16 g, yield = 71% in a ratio of **8l**/**9l** 83/17 in favor of the *trans* isomer **8l**.

8l. ^1H NMR (400 MHz, C_6D_6) δ 7.83 (d, J = 8.7 Hz, 2H), 6.84 (d, J = 8.7 Hz, 2H), 4.87 (t, J = 5.4 Hz, 1H), 3.99 (m, 1H), 4.09–3.92 (q, J = 7.1 Hz, 2H), 2.82 (s, 3H), 2.56 (s, 3H), 1.95 (d, J = 5.4 Hz, 2H), 1.78 (dd, J = 14.6, 4.0 Hz, 1H), 1.58 (m, 1H), 1.51 (dd, J = 14.6, 5.3 Hz, 1H), 1.37–1.25 (m, 3H), 0.97 (t, J = 7.5 Hz, 3H), 0.81 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, C_6D_6) δ 155.9, 150.5, 146.8, 126.7, 123.2, 98.7, 61.2, 53.6, 52.0, 47.4, 47.0, 37.0, 36.4, 34.1, 20.6, 14.5, 14.1; HRMS-ESI ($\text{M} + \text{Na}$) m/z calcd for $\text{C}_{19}\text{H}_{28}\text{N}_2\text{O}_6\text{Na}$ 403.1845, found 403.1831.

(2*S*,6*R*)-Ethyl-4,4-dimethoxy-6-methyl-2-[(4-nitrophenyl)]piperidine-1-carboxylate **8m** and (2*R*,6*R*)-Ethyl-4,4-dimethoxy-6-methyl-2-[(4-nitrophenyl)]piperidine-1-carboxylate **9m**. Yellow oil, starting from 1.0 mmol of **7d**, 0.28g, yield = 80% in a ratio of **8m**/**9m** 89/11 in favor of the *trans* isomer **8m**.

8m. ^1H NMR (400 MHz, C_6D_6) δ 7.87 (d, J = 8.8 Hz, 2H), 6.84 (d, J = 8.8 Hz, 2H), 4.98 (t, J = 5.3 Hz, 1H), 4.26 (m, 1H), 4.09–3.92 (m, 2H), 2.87 (s, 3H), 2.58 (s, 3H), 2.03 (d, J = 5.3 Hz, 2H), 1.67 (dd, J = 14.4, 3.5 Hz, 1H), 1.61 (dd, J = 14.4, 5.5 Hz, 1H), 1.37 (d, J = 6.7 Hz, 3H), 0.86 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, C_6D_6) δ 155.7, 150.8, 146.8, 123.3, 126.6, 98.2, 61.2, 53.7, 47.6, 47.2, 47.0, 37.6, 36.7, 20.6, 14.4.

9m. ^1H NMR (400 MHz, C_6D_6) δ 7.84 (d, J = 8.8 Hz, 2H), 7.06 (d, J = 8.8 Hz, 2H), 5.19 (dd, J = 6.9, 6.6 Hz, 1H), 4.43 (m, 1H), 4.09–3.92 (m, 2H), 2.98 (s, 3H), 2.79 (s, 3H), 2.01 (dd, J = 14.1, 6.9 Hz, 1H), 1.94 (dd, J = 14.1, 6.6 Hz, 1H), 1.79 (dd, J = 14.2, 6.7 Hz, 1H), 1.43 (dd, J = 14.2, 5.9 Hz, 1H), 1.16 (d, J = 6.7 Hz, 3H), 0.87 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, C_6D_6) δ 155.7, 151.9, 146.8, 123.8, 126.9, 98.6, 61.5, 54.0, 48.2, 47.2, 47.0, 36.9, 34.0, 23.0, 14.4; HRMS ($\text{M} + \text{H}$) $^+$ ion by direct probe calcd for $\text{C}_{17}\text{H}_{25}\text{N}_2\text{O}_6$ 353.1713, found 353.1699.

(2*S*,6*R*)-Ethyl-4,4-dimethoxy-6-methyl-2-[(3-nitrophenyl)]piperidine-1-carboxylate **8n** and (2*R*,6*R*)-Ethyl-4,4-dimethoxy-6-methyl-2-[(3-nitrophenyl)]piperidine-1-carboxylate **9n**. Yellow oil, starting from 1.0 mmol of **7c**, 0.25 g, yield = 71% in a ratio of **8n**/**9n** 40/60 in favor of the *cis* isomer **9n**.

8n. ^1H NMR (400 MHz, C_6D_6) δ 8.21 (m, 1H), 7.87 (d, J = 8.0 Hz, 1H), 7.24 (d, J = 7.8 Hz, 1H), 6.91 (dd, J = 8.0, 7.8 Hz, 1H), 4.98 (dd, J = 5.3, 4.5 Hz, 1H), 4.26 (m, 1H), 4.18–4.10 (m, 2H), 2.96 (s, 3H), 2.71 (s, 3H), 2.18 (dd, J = 14.4, 4.5 Hz, 1H), 2.13 (dd, J = 14.4, 5.3 Hz, 1H), 1.67 (dd, J = 14.4, 4.0 Hz, 1H), 1.73 (dd, J = 14.4, 4.8 Hz, 1H), 1.47 (d, J = 6.8 Hz, 3H), 1.02 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101

MHz, C_6D_6) δ 155.8, 148.7, 145.6, 131.6, 128.8, 121.3, 121.1, 98.2, 61.2, 53.5, 47.5, 47.2, 47.0, 37.4, 36.8, 20.6, 14.5.

9n. ^1H NMR (400 MHz, C_6D_6) δ 8.40 (m, 1H), 7.87 (d, J = 8.0 Hz, 1H), 7.45 (d, J = 7.8 Hz, 1H), 6.91 (dd, J = 8.0, 7.8 Hz, 1H), 5.40 (dd, J = 6.8, 6.6 Hz, 1H), 4.55 (qd, J = 6.8, 5.8 Hz, 1H), 4.13–4.04 (m, 2H), 3.02 (s, 3H), 2.90 (s, 3H), 2.19 (dd, J = 14.6, 6.8 Hz, 1H), 2.06 (ddd, J = 14.6, 6.6, 0.7 Hz, 1H), 1.90 (ddd, J = 14.1, 6.8, 0.7 Hz, 1H), 1.55 (dd, J = 14.4, 5.8 Hz, 1H), 1.29 (d, J = 6.8 Hz, 3H), 0.99 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, C_6D_6) δ 156.3, 148.7, 146.8, 132.2, 129.0, 121.5, 121.4, 98.6, 61.5, 54.0, 47.6, 47.6, 47.2, 37.4, 36.7, 23.0, 14.5; HRMS ($\text{M} + \text{H}$) $^+$ ion by direct probe calcd for $\text{C}_{17}\text{H}_{25}\text{N}_2\text{O}_6$ 353.1713, found 353.1710.

(2*S*,6*R*)-Ethyl-4,4-dimethoxy-6-methyl-2-[(2-nitrophenyl)]piperidine-1-carboxylate **8o** and (2*R*,6*R*)-Ethyl-4,4-dimethoxy-6-methyl-2-[(2-nitrophenyl)]piperidine-1-carboxylate **9o**. Yellow oil, starting from 1.0 mmol of **7b**, 0.23 g, yield = 67% in a ratio of **8o**/**9o** 60/40 in favor of the *trans* isomer **8o**.

8o. ^1H NMR (400 MHz, C_6D_6) δ 7.51 (dd, J = 8.0, 1.2 Hz, 1H), 7.50 (d, J = 7.8 Hz, 1H), 7.07 (td, J = 8.0, 1.0 Hz, 1H), 6.77 (ddd, J = 8.0, 7.8, 1.2 Hz, 1H), 5.59 (dd, J = 12.4, 5.3 Hz, 1H), 4.72 (qtd, J = 6.8, 4.5 Hz, 1H), 3.91–3.71 (m, 2H), 3.2 (s, 3H), 3.01 (s, 3H), 2.18 (ddd, J = 14.4, 5.3, 1.2 Hz, 1H), 2.80 (ddd, J = 14.4, 6.8, 1.2 Hz, 1H), 1.95 (dd, J = 14.4, 12.4 Hz, 1H), 1.71 (dd, J = 14.4, 4.5 Hz, 1H), 1.52 (d, J = 6.8 Hz, 3H), 0.74 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, C_6D_6) δ 156.1, 149.6, 141.5, 132.6, 127.6, 126.9, 124.5, 98.4, 61.2, 52.2, 47.9, 47.7, 47.0, 37.9, 36.9, 24.0, 13.9.

9o. ^1H NMR (400 MHz, C_6D_6) δ 7.55 (dd, J = 8.0, 1.2 Hz, 1H), 7.32 (d, J = 7.8 Hz, 1H), 7.03 (td, J = 8.0, 1.0 Hz, 1H), 6.76 (ddd, J = 8.0, 7.8, 1.2 Hz, 1H), 5.94 (dd, J = 6.6, 5.3 Hz, 1H), 4.55 (m, 1H), 4.02–3.84 (m, 2H), 3.00 (s, 3H), 2.79 (s, 3H), 2.40 (dd, J = 14.4, 5.3 Hz, 1H), 2.21 (dd, J = 14.4, 6.6 Hz, 1H), 1.86 (dd, J = 14.4, 5.5 Hz, 1H), 1.80 (dd, J = 14.4, 3.3 Hz, 1H), 1.47 (d, J = 6.8 Hz, 3H), 0.89 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, C_6D_6) δ 156.0, 149.2, 139.3, 132.0, 127.8, 127.0, 123.8, 98.1, 61.1, 50.6, 48.5, 47.7, 47.4, 38.8, 37.4, 20.2, 14.3; HRMS ($\text{M} + \text{H}$) $^+$ ion by direct probe calcd for $\text{C}_{17}\text{H}_{25}\text{N}_2\text{O}_6$ 353.1713, found 353.1711.

(2*S*,6*R*)-ethyl-2-(4-bromophenyl)-4,4-dimethoxy-6-methylpiperidine-1-carboxylate **8p** and (2*R*,6*R*)-ethyl-2-(4-bromophenyl)-4,4-dimethoxy-6-methylpiperidine-1-carboxylate **9p**. Yellow oil, starting from 0.9 mmol of **7g**, 0.25 g, yield = 73% in a ratio of **8p**/**9p** 33/67 in favor of the *cis* isomer **9p**.

8p. ^1H NMR (400 MHz, C_6D_6) δ 7.13 (d, J = 8.3 Hz, 2H), 6.68 (d, J = 8.3 Hz, 2H), 4.91 (dd, J = 5.3, 4.5 Hz, 1H), 4.15 (m, 1H), 3.91 (qd, J = 7.1 Hz, 2H), 2.75 (s, 3H), 2.49 (s, 3H), 2.10 (dd, J = 14.4, 4.5 Hz, 1H), 2.03 (dd, J = 14.4, 5.3 Hz, 1H), 1.82 (dd, J = 14.4, 5.3 Hz, 1H), 1.78 (dd, J = 14.4, 3.8 Hz, 1H), 1.40 (d, J = 6.8 Hz, 3H), 0.89 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, C_6D_6) δ 155.9, 142.5, 131.3, 128.0, 120.1, 98.5, 61.0, 53.5, 47.7, 47.5, 47.4, 37.5, 36.7, 20.6, 14.6.

9p. ^1H NMR (400 MHz, C_6D_6) δ 7.12 (d, J = 8.6 Hz, 2H), 6.92 (d, J = 8.6 Hz, 2H), 5.13 (t, J = 6.8 Hz, 1H), 4.32 (sex, J = 6.6 Hz, 1H), 3.88 (qd, J = 7.1 Hz, 2H), 2.79 (s, 3H), 2.69 (s, 3H), 2.09 (dd, J = 14.6, 6.8 Hz, 1H), 2.01 (ddd, J = 14.6, 6.8, 0.7 Hz, 1H), 1.82 (ddd, J = 14.1, 6.6, 0.7 Hz, 1H), 1.49 (dd, J = 14.1, 6.6 Hz, 1H), 1.18 (d, J = 6.6 Hz, 3H), 0.87 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, C_6D_6) δ 156.5, 143.8, 131.5, 128.3, 120.5, 98.8, 61.3, 53.8, 47.2, 47.0, 37.6, 36.9, 23.0, 14.5; HRMS-ESI ($\text{M} + \text{Na}$) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{NO}_4\text{BrNa}$ 408.0786, found 408.0794.

(2*S*,6*R*)-ethyl-2-(4-bromophenyl)-4,4-dimethoxy-6-methylpiperidine-1-carboxylate **8q** and (2*R*,6*R*)-ethyl-2-(4-bromophenyl)-4,4-dimethoxy-6-methylpiperidine-1-carboxylate **9q**. Yellow oil, starting from 0.9 mmol of **7f**, 0.26 g, yield = 75% in a ratio of **8q**/**9q** 29/71 in favor of the *cis* isomer **9q**.

8q. ^1H NMR (400 MHz, C_6D_6) δ 7.51 (dd, J = 7.7, 1.2 Hz, 1H), 7.47 (dd, J = 7.7, 0.7 Hz, 1H), 7.09 (t, J = 7.7 Hz, 1H), 6.79 (t, J = 7.7 Hz, 1H), 5.58 (dd, J = 12.4, 5.1 Hz, 1H), 4.85 (qtd, J = 7.0, 3.7 Hz, 1H), 3.91–4.08 (m, 2H), 3.15 (s, 3H), 2.97 (s, 3H), 2.57 (dd, J = 14.3, 5.1 Hz, 1H), 2.11 (dd, J = 14.1, 7.5 Hz, 1H), 1.93 (dd, J = 14.3, 12.4 Hz, 1H), 1.72 (dd, J = 14.1, 3.7 Hz, 1H), 1.40 (d, J = 7.0 Hz, 3H), 0.83 (t, J = 7.1 Hz, 3H). ^{13}C NMR (101 MHz, C_6D_6) δ 156.8, 133.0, 129.7,

128.3, 127.9, 126.8, 98.6, 61.1, 56.0, 47.5, 47.4, 47.3, 39.0, 37.4, 24.1, 14.1.

9q. $^1\text{H NMR}$ (400 MHz, C_6D_6) δ 7.77 (d, $J = 8.2$ Hz, 1H), 7.46 (dd, $J = 7.7, 0.7$ Hz, 1H), 7.46 (t, $J = 7.7$ Hz, 1H), 6.76 (m, 1H), 5.56 (dd, $J = 5.8, 5.4$ Hz, 1H), 4.69 (m, 1H), 3.90 (m, 2H), 3.03 (s, 3H), 2.80 (s, 3H), 2.62 (dd, $J = 14.5, 5.4$ Hz, 1H), 2.33 (dd, $J = 14.5, 5.8$ Hz, 1H), 2.05 (dd, $J = 14.6, 6.2$ Hz, 1H), 1.89 (m, 1H), 1.57 (d, $J = 6.9$ Hz, 3H), 0.91 (t, $J = 6.9$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, C_6D_6) δ 156.3, 133.1, 128.6, 128.3, 127.9, 127.2, 98.5, 60.9, 54.6, 48.2, 48.0, 47.0, 37.2, 36.2, 20.7, 14.4; HRMS-ESI ($\text{M} + \text{Na}$) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{NO}_4\text{BrNa}$ 408.0786, found 408.0790

(2*S*,6*R*)-ethyl-4,4-dimethoxy-6-methyl-2-[(3-nitro-5-chlorophenyl)]piperidine-1-carboxylate **8r** and (2*S*,6*R*)-ethyl-4,4-dimethoxy-6-methyl-2-[(3-nitro-5-chlorophenyl)]piperidine-1-carboxylate **9r**. Yellow oil, starting from 1.2 mmol of **7h**, 0.35 g; yield = 76% in a ratio of **8r/9r** 56/43 in favor of the *trans* isomer **8r**.

8r. $^1\text{H NMR}$ (400 MHz, C_6D_6) δ 7.60 (d, $J = 8.6$ Hz, 1H), 7.56 (d, $J = 2.3$ Hz, 1H), 6.60 (dd, $J = 8.6, 2.3$ Hz, 1H), 5.47 (dd, $J = 12.1, 5.3$ Hz, 1H), 4.49 (qtd, $J = 6.8, 4.8$ Hz, 1H), 3.77–3.58 (m, 2H), 3.03 (s, 3H), 2.87 (s, 3H), 2.59 (ddd, $J = 14.4, 5.3, 1.5$ Hz, 1H), 1.89 (ddd, $J = 14.4, 6.8, 1.5$ Hz, 1H), 1.75 (dd, $J = 14.4, 12.1$ Hz, 1H), 1.48 (dd, $J = 14.4, 4.8$ Hz, 1H), 1.32 (d, $J = 6.8$ Hz, 3H), 0.63 (t, $J = 7.1$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, C_6D_6) δ 156.0, 147.7, 143.8, 139.0, 127.9, 127.2, 125.6, 98.2, 61.4, 52.5, 47.9, 48.5, 47.5, 38.6, 37.5, 19.2, 13.9.

9r. $^1\text{H NMR}$ (400 MHz, C_6D_6) δ 7.44 (d, $J = 2.3$ Hz, 1H), 6.6 (d, $J = 8.6$ Hz, 1H), 6.59 (dd, $J = 8.6, 2.3$ Hz, 1H), 5.70 (dd, $J = 6.8, 5.3$ Hz, 1H), 4.30 (m, 1H), 3.90–3.75 (m, 2H), 2.86 (s, 3H), 2.68 (s, 3H), 2.29 (dd, $J = 14.4, 5.3$ Hz, 1H), 1.96 (dd, $J = 14.4, 6.8$ Hz, 1H), 1.68 (dd, $J = 14.4, 5.5$ Hz, 1H), 1.64 (dd, $J = 14.4, 3.3$ Hz, 1H), 1.28 (d, $J = 6.8$ Hz, 3H), 0.78 (t, $J = 7.1$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, C_6D_6) δ 156.0, 147.7, 141.7, 138.6, 129.7, 127.2, 126.2, 98.0, 61.3, 50.5, 47.7, 47.5, 47.1, 37.3, 36.9, 24.0, 14.3; HRMS ($\text{M} + \text{H}$)⁺ ion by direct probe calcd for $\text{C}_{17}\text{H}_{24}\text{ClN}_2\text{O}_6$ 387.1323, found 387.1325.

(2*S*,6*R*)-ethyl-2-(*E*)-3'-ethoxy-3'-oxoprop-1'-enyl)-4,4-dimethoxy-6-methylpiperidine-1-carboxylate **8s**. Yellow oil, starting from 0.7 mmol of **7j**, 0.21 g, yield = 93% in a ratio of **8s/9s**: 90/10 in favor of the *trans* isomer **8s**.

8s. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.89 (dd, $J = 15.7, 5.3$ Hz, 1H), 5.71 (dd, $J = 15.7, 2.0$ Hz, 1H), 4.60 (tdd, $J = 5.3, 4.2, 2.0$ Hz, 1H), 4.14–4.02 (m, 5H), 3.12 (s, 3H), 3.06 (s, 3H), 2.13 (dd, $J = 14.2, 5.3$ Hz, 1H), 2.06 (dd, $J = 14.2, 4.2$ Hz, 1H), 1.92 (dd, $J = 14.4, 4.5$ Hz, 1H), 1.83 (dd, $J = 14.4, 3.3$ Hz, 1H), 1.24–1.17 (m, 9H); $^{13}\text{C NMR}$ (101 MHz, C_6D_6) δ 166.1, 155.4, 149.4, 141.2, 98.2, 61.2, 60.1, 51.7, 47.3, 47.2, 47.0, 37.1, 36.8, 20.7, 14.7, 14.2; HRMS-ESI ($\text{M} + \text{Na}$) m/z calcd for $\text{C}_{16}\text{H}_{27}\text{NO}_6\text{Na}$ 352.1736, found 352.1729.

General Procedure for the Synthesis of Piperidines 10/11. (7*R*,9*S*)-Ethyl-7-methyl-9-phenyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **10c** and (7*S*,9*S*)-Ethyl-7-methyl-9-phenyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **11c**. To a solution of compound **7w** (100 mg, 0.38 mmol, 1 equiv) were added successively trimethyl orthoformate (0.21 mL, 1.90 mmol, 5 equiv) and *p*-toluenesulphonic acid PTSA/ H_2O (1.5 mg, 0.08 mmol, 0.2 equiv). The reaction was followed by TLC, and after 0.5 h ethyl acetate was added to the crude mixture, quenched with a saturated solution of NaHCO_3 , and extracted twice with ethyl acetate. Then the organic layer was dried and concentrated under vacuum. After $^1\text{H NMR}$ spectroscopy for identifying the product and measuring the de, the crude mixture **8c/9c** (0.117 g, 0.38 mmol, ratio **8c/9c** 44/56) was engaged in reaction without any purification. To a solution of **8c/9c** in dry CH_2Cl_2 (5 mL) were added successively 1,2-ethanedithiol (161 mL, 1.9 mmol, 5 equiv) and $\text{BF}_3 \cdot \text{Et}_2\text{O}$ (243 mL, 1.9 mmol, 5 equiv) dropwise at 0 °C. After 2 h at room temperature, the solution was quenched dropwise with a solution of NaOH at 0 °C and extracted three times with ethyl acetate. Then the organic layer was dried over Na_2SO_4 and concentrated under vacuum before being purified by flash chromatography (eluent, cyclohexane to cyclohexane/ EtOAc 9:1) to give a mixture of both isomer **10c** and **11c** (0.088 g, 68%), ratio **10c/11c** 44/56). HRMS-ESI ($\text{M} + \text{Na}$) m/z calcd for $\text{C}_{17}\text{H}_{23}\text{NO}_2\text{S}_2\text{Na}$ 360.1068, found 360.1069.

10c. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.36–7.12 (m, 5H), 5.26 (dd, $J = 4.8, 5.0$ Hz, 1H), 4.41 (m, 1H), 3.97 (q, $J = 7.1$ Hz, 2H), 3.14–3.01 (m, 4H), 2.80 (dd, $J = 5.0, 14.9$ Hz, 1H), 2.74 (dd, $J = 4.8, 14.9$ Hz, 1H), 2.51 (dd, $J = 5.0, 14.9$ Hz, 1H), 2.22 (dd, $J = 4.3, 14.9$ Hz, 1H), 1.43 (d, $J = 7.1$ Hz, 3H), 1.03 (t, $J = 7.1$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 156.2, 141.2, 128.2, 126.7, 125.9, 61.1, 61.0, 55.2, 48.6, 46.3, 46.0, 39.3, 39.2, 20.5, 14.5

11c. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.36–7.12 (m, 5H), 5.26 (dd, $J = 7.8, 8.3$ Hz, 1H), 4.41 (tdd, $J = 7.0, 7.5, 6.8$ Hz, 1H), 4.03 (q, $J = 7.1$ Hz, 2H), 3.26–3.21 (m, 4H), 2.63 (ddd, $J = 7.8, 14.6, 1.8$ Hz, 1H), 2.57 (dd, $J = 8.3, 14.6$ Hz, 1H), 2.44 (ddd, $J = 7.0, 14.4, 1.8$ Hz, 1H), 2.03 (dd, $J = 7.5, 14.4$ Hz, 1H), 1.22 (d, $J = 6.8$ Hz, 3H), 1.06 (t, $J = 7.1$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 156.7, 143.7, 128.3, 126.6, 125.9, 62.4, 61.4, 55.7, 48.2, 46.2, 45.5, 39.2, 39.1, 23.5, 14.5.

(7*R*,9*S*)-7-Methyl-9-phenyl-1,4-dithia-8-azaspiro[4.5]decane **10a** and (7*R*,9*R*)-7-Methyl-9-phenyl-1,4-dithia-8-azaspiro[4.5]decane **11a**. Yellow oil, starting from 0.8 mmol of **7a**, 0.15 g, yield = 68% in a ratio of **10a/11a** 86/14 in favor of the *trans* isomer **10a**.

10a. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.38–7.25 (m, 5H), 4.18 (dd, $J = 8.8, 3.4$ Hz, 1H), 3.47 (m, 1H), 3.35–3.10 (m, 4H), 2.38 (ddd, $J = 13.7, 5.2$ Hz, 1H), 2.28 (ddd, $J = 13.4, 3.4, 1.6$ Hz, 1H), 2.18 (dd, $J = 13.3, 8.8$ Hz, 1H), 1.96 (ddd, $J = 13.7, 4.0, 1.6$ Hz, 1H), 1.52 (s, 1H), 1.30 (t, $J = 7.0$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 143.5, 128.4, 127.0, 126.7, 65.2, 53.8, 49.9, 48.1, 45.8, 39.6, 37.7, 20.8; HRMS ($\text{M} + \text{H}$)⁺ ion by direct probe calcd for $\text{C}_{14}\text{H}_{19}\text{NS}_2$ 266.1037, found 266.1042

11a. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.38–7.25 (m, 5H), 3.85 (dd, $J = 11.2, 2.3$ Hz, 1H), 3.30–3.20 (m, 4H), 2.95 (m, 1H), 2.15 (dt, $J = 13.1, 2.3$ Hz, 1H), 2.05 (dt, $J = 12.9, 2.3$ Hz, 1H), 1.95 (dd, $J = 13.3, 11.2$ Hz, 1H), 1.75 (dd, $J = 12.9, 11.0$ Hz, 1H), 1.08 (d, $J = 7.0$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 142.8, 127.4, 126.3, 125.8, 65.9, 59.9, 51.0, 49.4, 48.7, 38.2, 36.7, 21.3; HRMS ($\text{M} + \text{H}$)⁺ ion by direct probe calcd for $\text{C}_{14}\text{H}_{19}\text{NS}_2$ 266.1037, found 266.1040.

(7*S*,9*R*)-Ethyl-7-phenyl-9-propyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **10f** and (7*R*,9*R*)-Ethyl-7-phenyl-9-propyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **11f**. Yellow oil, starting from 1.0 mmol of **7q**, 0.22 g, yield = 60% (entry 6, Table 1) in a ratio of **10f/11f** $\geq 95/5$ in favor of the *trans* isomer **10f**.

10f. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.26–7.12 (m, 5H), 5.20 (dd, $J = 9.3, 7.3$ Hz, 1H), 4.29 (m, 1H), 4.02 (q, $J = 7.1$ Hz, 2H), 3.28–3.20 (m, 4H), 2.56 (ddd, $J = 14.4, 7.3, 1.8$ Hz, 1H), 2.53 (dd, $J = 14.4, 9.3$ Hz, 1H), 2.50 (ddd, $J = 14.1, 8.4, 1.8$ Hz, 1H), 2.11 (dd, $J = 14.1, 5.8$ Hz, 1H), 1.67 (m, 1H), 1.42–1.21 (m, 3 H), 1.04 (t, $J = 6.9$ Hz, 3H), 0.82 (t, $J = 7.1$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 157.0, 143.8, 128.2, 126.6, 125.9, 62.4, 61.4, 55.8, 52.1, 45.9, 44.2, 39.2, 39.1, 19.7, 14.4, 13.8.

11f. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.26–7.12 (m, 5H), 5.09 (dd, $J = 5.5, 5.2$ Hz, 1H), 4.04–3.95 (m, 3H), 3.17–3.03 (m, 4H), 2.75 (dd, $J = 14.6, 5.2$ Hz, 1H), 2.68 (dd, $J = 14.6, 5.5$ Hz, 1H), 2.36 (dd, $J = 14.8, 4.6$ Hz, 1H), 2.32 (dd, $J = 14.8, 4.8$ Hz, 1H), 1.85 (m, 1H), 1.42–1.21 (m, 3 H), 1.00 (t, $J = 7.1$ Hz, 3H), 0.91 (t, $J = 7.1$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, C_6D_6) δ 156.1, 141.9, 128.4, 126.8, 126.2, 62.3, 61.0, 55.8, 53.5, 45.9, 43.6, 40.7, 36.1, 20.3, 14.2, 13.9; HRMS-ESI ($\text{M} + \text{Na}$) m/z calcd for $\text{C}_{19}\text{H}_{27}\text{NO}_2\text{S}_2\text{Na}$ 388.1381, found 388.1377.

(7*R*,9*S*)-Ethyl-7-octyl-9-phenyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **10h** and (7*R*,9*R*)-Ethyl-7-octyl-9-phenyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **11h**. Yellow oil, starting from 0.8 mmol of **7s**, 0.23 g, yield = 65% (entry 8, Table 1) in a ratio of **10h/11h** 62/38 in favor of the *trans* isomer **10h**.

10h. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.25–7.12 (m, 5H), 5.20 (dd, $J = 9.1, 7.3$ Hz, 1H), 4.27 (m, 1H), 4.00 (q, $J = 7.1$ Hz, 2H), 3.28–3.20 (m, 4H), 2.56 (ddd, $J = 14.4, 7.3, 1.5$ Hz, 1H), 2.50 (dd, $J = 14.4, 9.3$ Hz, 1H), 2.49 (ddd, $J = 14.1, 8.8, 1.5$ Hz, 1H), 2.09 (dd, $J = 14.1, 5.5$ Hz, 1H), 1.67 (m, 1H), 1.38–1.11 (m, 15 H), 1.04 (t, $J = 7.1$ Hz, 3H), 0.80 (t, $J = 6.8$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 157.0, 143.8, 128.2, 126.6, 126.0, 62.5, 61.4, 55.8, 52.4, 45.9, 44.3, 39.2, 39.1, 38.0, 31.8, 29.5, 29.4, 26.5, 22.7, 14.5, 14.1.

11h. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.25–7.12 (m, 5H), 5.09 (t, $J = 5.1$ Hz, 1H), 4.15 (m, 1H), 3.96 (q, $J = 7.1$ Hz, 2H), 3.17–3.02 (m, 4H), 2.76 (dd, $J = 14.6, 5.1$ Hz, 1H), 2.69 (dd, $J = 14.6, 5.1$ Hz, 1H),

2.36 (dd, $J = 14.8, 5.1$ Hz, 1H), 2.32 (dd, $J = 14.8, 4.2$ Hz, 1H), 1.85 (m, 1H), 1.76–1.01 (m, 15 H), 1.08 (t, $J = 7.1$ Hz, 3H), 0.82 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 128.4, 126.8, 126.2, 62.3, 61.0, 55.8, 53.5, 45.9, 43.6, 40.7, 37.9, 36.1, 32.0, 29.4, 29.6, 26.3, 22.6, 14.6, 14.0; HRMS-ESI (M + Na) m/z calcd for $\text{C}_{24}\text{H}_{37}\text{NO}_2\text{S}_2\text{Na}$ 458.2163, found 458.2165.

(7*S*,9*S*)-Ethyl-7,9-diphenyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **10j**. Yellow oil, starting from 0.6 mmol of **7v**, 0.16 g, yield = 65% in a ratio of **10j/11j** $\geq 6/4$ in favor of the *trans* isomer **10j**. ^1H NMR (400 MHz, CDCl_3) δ 7.25–7.11 (m, 10H), 5.22 (dd, $J = 8.6, 6.3$ Hz, 2H), 3.91 (q, $J = 7.1$ Hz, 2H), 3.31–3.23 (m, 4H), 2.64 (dd, $J = 14.4, 8.6$ Hz, 2H), 2.52 (dd, $J = 14.4, 6.3$ Hz, 2H), 0.89 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 157.6, 143.6, 128.2, 126.8, 126.7, 62.4, 61.5, 57.5, 46.0, 39.5, 39.0, 14.1; HRMS-ESI (M + Na) m/z calcd for $\text{C}_{22}\text{H}_{25}\text{NO}_2\text{S}_2\text{Na}$ 422.1224, found 422.1225.

(9*R*,7*S*)-Ethyl-9-methyl-7-(pyridin-3-yl)-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **10k**. Yellow oil, yield = 48% in a ratio of **10k/11k** 85/15 in favor of the *trans* isomer **10k**. ^1H NMR (400 MHz, CDCl_3) δ 8.46 (m, 1H), 8.42 (d, $J = 4.5$ Hz, 1H), 7.48 (d, $J = 8.0$ Hz, 1H), 7.21 (m, 1H), 5.13 (t, $J = 4.9$ Hz, 1H), 4.26 (m, 1H), 4.07–3.93 (m, 2H), 3.30–3.01 (m, 4H), 2.78 (dd, $J = 15.0, 4.9$ Hz, 1H), 2.69 (dd, $J = 15.0, 4.9$ Hz, 1H), 2.44 (dd, $J = 15.0, 5.1$ Hz, 1H), 2.24 (dd, $J = 15.1, 3.8$ Hz, 1H), 1.42 (d, $J = 6.9$ Hz, 3H), 1.05 (t, $J = 6.9$ Hz); ^{13}C NMR (101 MHz, CDCl_3) δ 155.0, 146.7, 146.6, 136.6, 122.1, 60.4, 59.8, 52.4, 47.7, 45.3, 44.9, 38.6, 38.0, 19.4, 13.4; HRMS-ESI (M + Na) m/z calcd for $\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}_2\text{S}_2\text{Na}$ 361.1020, found 361.1022.

(7*R*,9*R*)-Ethyl-7-methyl-9-propyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **10v** and (7*R*,9*S*)-Ethyl-7-methyl-9-propyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **11v**. Yellow oil, starting from 0.9 mmol of **7l**, 0.20 g, yield = 73% in a ratio of **10v/11v** 80/20 in favor of the *trans* isomer **10v**.

10v. ^1H NMR (400 MHz, CDCl_3) δ 4.12–3.96 (m, 2H), 3.93 (qdd, $J = 7.0, 5.1, 4.8$ Hz, 1H), 3.85 (m, 1H), 3.32–3.13 (m, 4H), 2.39 (dd, $J = 14.7, 4.8$ Hz, 1H), 2.35 (dd, $J = 14.9, 4.8$ Hz, 1H), 2.31 (dd, $J = 14.9, 4.3$ Hz, 1H), 2.17 (dd, $J = 14.7, 5.1$ Hz, 1H), 1.69 (m, 1H), 1.55 (m, 1H), 1.37 (m, 1H), 1.29 (d, $J = 7.0$ Hz, 3H), 1.25 (m, 1H), 1.18 (t, $J = 7.1$ Hz, 3H), 0.86 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 155.8, 61.7, 60.8, 52.4, 47.9, 46.7, 42.9, 39.4, 39.1, 35.5, 20.4, 19.8, 14.6, 13.8; HRMS (M + H) $^+$ ion by direct probe calcd for $\text{C}_{14}\text{H}_{26}\text{NO}_2\text{S}_2$ 304.1405, found 304.1394.

11v. ^1H NMR (400 MHz, CDCl_3) δ 4.34 (m, 1H), 4.26 (m, 1H), 4.09 (q, $J = 7.1$ Hz, 2H), 3.25–3.13 (m, 4H), 2.45 (ddd, $J = 13.9, 9.1, 1.8$ Hz, 1H), 2.35 (ddd, $J = 13.6, 8.1, 1.8$ Hz, 1H), 1.98 (dd, $J = 13.9, 7.5$ Hz, 1H), 1.95 (dd, $J = 13.6, 7.3$ Hz, 1H), 2.18–1.45 (m, 2H), 1.55 (m, 2H), 1.37–1.23 (m, 2H), 1.21 (d, $J = 6.8$ Hz, 3H), 1.98 (t, $J = 7.1$ Hz, 3H), 0.84 (t, $J = 7.5$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 156.1, 61.9, 61.2, 52.4, 50.5, 47.6, 45.5, 39.4, 39.0, 35.5, 22.5, 19.6, 14.7, 13.9; HRMS (M + H) $^+$ ion by direct probe calcd for $\text{C}_{14}\text{H}_{26}\text{NO}_2\text{S}_2$ 304.1405, found 304.1401.

(7*R*,9*R*)-Ethyl-7-methyl-9-octyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **10t** and (7*R*,9*S*)-Ethyl-7-methyl-9-octyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **11t**. Yellow oil, starting from 1.0 mmol of **7m**, 0.23 g, yield = 62%; entry 5 Table S, in a ratio of **10t/11t** 80/20 in favor of the *trans* isomer **10t**.

10t. ^1H NMR (400 MHz, CDCl_3) δ 4.06 (qd, $J = 7.1, 3.5$ Hz, 2H), 3.92 (qdd, $J = 6.8, 5.3, 4.8$ Hz, 1H), 3.83 (m, 1H), 3.32–3.15 (m, 4H), 2.40 (dd, $J = 14.6, 4.8$ Hz, 1H), 2.36 (dd, $J = 15.1, 5.0$ Hz, 1H), 2.30 (dd, $J = 15.1, 4.3$ Hz, 1H), 2.17 (dd, $J = 14.6, 5.3$ Hz, 1H), 1.70 (m, 1H), 1.59 (m, 1H), 1.37 (m, 1H), 1.31 (d, $J = 6.8$ Hz, 3H), 1.16–1.29 (m, 14H), 0.81 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 155.7, 61.6, 60.7, 52.6, 47.9, 46.6, 42.7, 39.3, 39.1, 33.2, 31.8, 29.4, 29.3, 29.2, 26.4, 22.6, 20.3, 14.6, 14.0; HRMS-ESI (M + Na) m/z calcd for $\text{C}_{19}\text{H}_{35}\text{NO}_2\text{S}_2\text{Na}$ 396.2007, found 396.2005

11t. ^1H NMR (400 MHz, CDCl_3) δ 4.34 (sex, $J = 7.1$ Hz, 1H), 4.24 (m, 1H), 4.10 (qd, $J = 7.1, 4.5$ Hz, 2H), 3.24–3.19 (m, 4H), 2.44 (ddd, $J = 13.9, 4.5, 2.0$ Hz, 1H), 2.35 (ddd, $J = 7.1, 13.9, 2.0$ Hz, 1H), 1.99 (dd, $J = 13.9, 7.1$ Hz, 1H), 1.96 (dd, $J = 13.9, 7.1$ Hz, 1H), 1.99–1.47 (m, 2H), 1.24–1.17 (m, 18H), 0.84 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 156.1, 62.9, 61.2, 51.0, 47.5, 43.9, 38.9, 38.7, 38.3, 31.8, 29.5, 29.4, 29.2, 26.4, 22.5, 14.7, 14.1; HRMS-

ESI (M + Na) m/z calcd for $\text{C}_{19}\text{H}_{35}\text{NO}_2\text{S}_2\text{Na}$ 396.2007, found 396.2003

(7*R*,9*R*)-Ethyl-7-octyl-9-propyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **10u** and (7*R*,9*S*)-Ethyl-7-octyl-9-propyl-1,4-dithia-8-azaspiro[4.5]decane-8-carboxylate **11u**. Yellow oil, starting from 0.6 mmol of **7u**, 0.16 g, yield = 68%; entry 1 Table S; in a ratio of **10u/11u** 80/20 in favor of the *trans* isomer **10u**.

10u. ^1H NMR (400 MHz, CDCl_3) δ 4.06 (qd, $J = 7.1, 2.7$ Hz, 2H), 3.74 (m, 2H), 3.30–3.15 (m, 4H), 2.62 (ddd, $J = 14.6, 4.8, 1.5$ Hz, 2H), 2.19 (dd, $J = 14.6, 5.6$ Hz, 2H), 1.87–1.73 and 1.58–1.45 (2 * m, 4H), 1.37–1.13 (m, 17H), 0.86 (t, $J = 7.3$ Hz, 3H), 0.86 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 156.1, 62.9, 60.8, 53.0, 52.7, 44.0, 43.9, 39.1, 39.0, 35.2, 33.0, 31.8, 29.5, 29.4, 29.3, 26.6, 22.6, 19.8, 14.6, 14.1, 13.9; HRMS-ESI (M + Na) m/z calcd for $\text{C}_{21}\text{H}_{39}\text{NO}_2\text{S}_2\text{Na}$ 424.2320, found 424.2327

11u. ^1H NMR (400 MHz, CDCl_3) δ 4.29–4.19 (m, 2H), 4.09 (q, $J = 7.1$ Hz, 2H), 3.20 (s, 4H), 2.41 (dd, $J = 13.9, 8.6$ Hz, 2H), 1.94 (dd, $J = 13.9, 6.1$ Hz, 2H), 1.62–1.56 and 1.49–1.40 (2 * m, 4H), 1.32–1.18 (m, 17H), 0.85 (t, $J = 7.6$ Hz, 3H), 0.81 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 156.4, 63.1, 61.2, 51.4, 51.1, 44.1, 44.0, 40.1, 39.1, 38.6, 37.9, 31.8, 29.5, 29.4, 29.2, 26.4, 22.6, 19.6, 14.7, 14.1, 13.9; HRMS-ESI (M + Na) m/z calcd for $\text{C}_{21}\text{H}_{39}\text{NO}_2\text{S}_2\text{Na}$ 424.2320, found 424.2323.

Synthesis of (–)-Solenopsine A from 7p. (7*R*,9*R*)-Benzyl-7-methyl-9-undecyl-1,4-dioxo-8-azaspiro[4.5]decane-8-carboxylate **8w**. To a solution of compound **7p** (400 mg, 1.07 mmol, 1 equiv) were added successively trimethyl orthoformate (0.587 mL, 5.36 mmol, 5 equiv), ethylene glycol (0.300 mL, 5.36 mmol 5 equiv), and *p*-toluenesulphonic acid (0.04 g, 0.21 mmol 0.2 equiv). The reaction is followed by TLC, and after 1 h ethyl acetate was added to the crude mixture, which was quenched with a saturated solution of NaHCO_3 and extracted twice with ethyl acetate. Then the organic layer was dried over Na_2SO_4 and concentrated under vacuum. The crude residue was purified by flash chromatography (eluent, cyclohexane/EtOAc 95:5 to cyclohexane/EtOAc 85:15), to afford *trans* **8w** (0.32 g, 67%): ^1H NMR (400 MHz, CDCl_3) δ 7.40–7.27 (m, 5H), 5.17 (d, $J = 12.4$ Hz, 1H), 5.09 (d, $J = 12.4$ Hz, 1H), 4.10 (m, 1H), 4.00–3.91 (m, 3H), 3.90–3.81 (m, 2H), 2.14 (dd, $J = 14.8, 5.6$ Hz, 1H), 2.05 (dd, $J = 15.2, 5.6$ Hz), 1.97 (dd, $J = 14.8, 3.8$ Hz, 1H), 1.81 (dd, $J = 15.2, 4.1$ Hz, 1H), 1.76–1.58 (m, 2H), 1.34 (d, $J = 7.1$ Hz, 3H), 1.37–1.22 (m, 16H), 0.88 (t, $J = 6.8$ Hz, 1H); ^{13}C NMR (101 MHz, CDCl_3) δ 155.6, 137.1, 128.6, 128.0, 106.7, 66.9, 64.0, 63.9, 51.5, 46.6, 39.6, 35.9, 34.2, 32.1, 29.8, 29.7, 29.5, 26.8, 21.0, 14.3; HRMS-ESI (M + Na) m/z calcd for $\text{C}_{27}\text{H}_{44}\text{NO}_4$ 446.3270 found 446.3261.

(7*R*,9*R*)-7-Methyl-9-undecyl-1,4-dithia-8-azaspiro[4.5]decane **10w**. To a solution of **8w** (300 mg, 0.64 mmol, 1 equiv) in dry CH_2Cl_2 (5 mL) were added successively 1,2-ethanedithiol (0.268 mL, 3.2 mmol, 5 equiv) and $\text{BF}_3\cdot\text{Et}_2\text{O}$ (0.406 mL, 3.2 mmol, 5 equiv) dropwise at 0 °C. After 24 h at room temperature, the solution was quenched with a solution of NaOH at 0 °C and extracted three times with CH_2Cl_2 . Then the organic layer was dried and concentrated under vacuum before purification by flash chromatography (eluent, cyclohexane to cyclohexane/EtOAc 70:30) to give *trans* **10w** (0.175 g, 76%) as a yellow oil: ^1H NMR (400 MHz, CDCl_3) δ 3.32–3.27 (m, 4H), 3.20 (m, 1H), 3.02 (m, 1H), 2.26 (dd, $J = 13.4, 3.9$ Hz, 1H), 2.22 (dd, $J = 13.1, 2.6$ Hz), 1.92 (dd, $J = 13.4, 5.6$ Hz, 1H), 1.82 (dd, $J = 13.1, 6.7$ Hz, 1H), 1.57–1.51 (m, 2H), 1.35–1.20 (m, 16H), 1.16 (d, $J = 6.7$ Hz, 3H), 0.88 (t, $J = 6.8$ Hz, 1H); ^{13}C NMR (101 MHz, CDCl_3) δ 65.3, 51.6, 49.2, 46.2, 46.0, 39.1, 38.7, 34.6, 32.1, 29.8, 29.7, 29.5, 26.9, 22.8, 21.0, 14.3; HRMS-ESI (M + Na) m/z calcd for $\text{C}_{19}\text{H}_{37}\text{NNaS}_2$ 366.2265 found 366.2257.

(2*R*,6*R*)-tert-Butyl-2-methyl-6-undecylpiperidine-1-carboxylate **12**. To a stirred solution of dithioacetal **10w** (0.140 g, 0.44 mmol, 1 equiv) in THF (5 mL) were added successively di-tert-butyl dicarbonate (0.191 g, 0.88 mmol, 2 equiv) and DMAP (25 mg, 0.02 mmol, 0.05 equiv) at 0 °C. After 1 h at room temperature the resulting solution mixture was washed with a solution of NH_4Cl and extracted with ethyl acetate. The organic layer was dried over Na_2SO_4 and concentrated under vacuum. The resulting oil was directly engaged in the following steps without further purification. To a solution of the

crude oil in ethanol (5 mL) was added freshly prepared W_2 Raney nickel (ca. 1 g). The resulting suspension was heated at reflux for 2 h and then cooled to room temperature. The suspension was then filtered through Celite, and the filtrate was concentrated under reduced pressure. The residue was dissolved in 1 M aqueous NaOH and extracted with dichloromethane. The combined organic extracts were washed with brine and dried over Na_2SO_4 . Evaporation of the solvent followed by column chromatography (eluent, cyclohexane to cyclohexane/EtOAc 80:20) gave compound **12** (100 mg, 70%) as a colorless oil. Spectral data are identical with those reported.⁴⁸

(-)-*Solenopsine A*. Trifluoroacetic acid (1 mL) was added to a solution of **12** (80 mg, 0.22 mmol) in CH_2Cl_2 (1 mL), and the reaction mixture was stirred at room temperature for 2 h. The mixture was evaporated, and the residue was basified with 2 N NaOH. The solution was extracted with CH_2Cl_2 three times. The extracts were dried with Na_2SO_4 and evaporated. The residue was purified with chromatography (eluent, $CHCl_3/MeOH$ 5:1) to yield (-)-solenopsine A (50 mg, 87%) as an oil. $[\alpha]_D = -1.21$ (c 0.94, CH_3OH), lit.⁴⁹ $[\alpha]_D = -1.30$ (c 1.30, CH_3OH). Spectral data are identical with those reported.⁴⁸

Synthesis of 241D from 7o. (2*R*,6*S*)-Benzyl-4,4-dimethoxy-2-methyl-6-nonylpiperidine-1-carboxylate **9'x**. To compound **7o** (0.25 g, 0.67 mmol, 1 equiv) were added successively trimethyl orthoformate (0.367 mL, 3.35 mmol, 5 equiv) and *p*-toluenesulphonic acid (127 mg, 0.67 mmol, 1 equiv). After 1 h, ethyl acetate was added to the crude mixture, which was quenched with a saturated solution of $NaHCO_3$ and extracted twice with ethyl acetate. Then the organic layer was dried and concentrated under vacuum. The crude oil mixture was separated by flash chromatography (eluent, cyclohexane/EtOAc 95:5 to cyclohexane/EtOAc 85:15) to afford pure *cis* isomer **9'x** as an oil (172 mg, 61%): 1H NMR (400 MHz, $CDCl_3$) δ 7.28–7.19 (m, 5H), 5.09 (d, $J = 12.6$ Hz, 1H), 5.05 (d, $J = 12.6$ Hz, 1H), 4.37 (quintd, $J = 7.1, 3.7$ Hz, 1H), 4.21 (qd, $J = 7.1, 2.1$ Hz, 1H), 3.11 (s, 6H), 1.87 (dt, $J = 14.0, 2.1$ Hz, 1H), 1.81 (ddd, $J = 13.7, 3.7, 2.1$ Hz, 1H), 1.73 (dd, $J = 14.0, 7.2$ Hz, 1H), 1.69 (dd, $J = 13.7, 7.7$ Hz, 1H), 1.61 (m, 2H), 1.23 (d, $J = 7.1$ Hz, 3H), 1.22–1.17 (m, 12H), 0.81 (t, $J = 6.8$ Hz, 1H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 155.6, 137.0, 128.7, 128.4, 127.9, 98.7, 67.0, 50.7, 47.9, 47.4, 46.3, 36.2, 35.7, 33.6, 31.9, 29.7, 29.6, 29.3, 27.2, 22.7, 21.5, 14.1; HRMS-ESI (M + Na) m/z calcd for $C_{25}H_{41}NO_4Na$ 442.2933 found 442.2948.

(2*R*,6*S*)-Benzyl-2-methyl-6-nonyl-4-oxopiperidine-1-carboxylate **13**. To a solution of **9'x** (0.150 g, 0.35 mmol, 1 equiv) in CH_2Cl_2 (0.5 mL) was added slowly TFA/ H_2O (1:1, 0.5 mL) at room temperature. After 1 h the mixture was quenched with NaOH (1 M) and extracted twice with CH_2Cl_2 . Then the organic layer was dried and concentrated under vacuum. The yellow oil obtained was filtered through a pad of silica and washed with ethyl acetate to furnish after evaporation of the solvent compound **13** as a yellow oil in a quantitative yield: 1H NMR (400 MHz, $CDCl_3$) δ 7.31–7.21 (m, 5H), 5.12 (d, $J = 12.4$ Hz, 1H), 5.08 (d, $J = 12.4$ Hz, 1H), 4.72 (m, 1H), 4.59 (m, 1H), 2.65 (dd, $J = 14.9, 7.7$ Hz, 1H), 2.60 (d, $J = 14.6, 7.5$ Hz, 1H), 2.27 (ddd, $J = 14.9, 3.7, 1.6$ Hz, 1H), 2.23 (ddd, $J = 14.6, 4.2, 1.6$ Hz, 1H), 1.59–1.35 (m, 2H), 1.22 (d, $J = 6.9$ Hz, 3H), 1.26–1.10 (m, 12H), 0.81 (t, $J = 6.9$ Hz, 1H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 208.1, 155.6, 136.5, 128.5, 128.1, 128.0, 67.5, 53.1, 48.8, 45.5, 43.7, 36.9, 31.9, 29.5, 29.3, 29.3, 26.3, 22.7, 21.5, 14.1; HRMS-ESI (M + Na) m/z calcd for $C_{23}H_{35}NO_3Na$ 396.2515 found 396.2511

(2*R*,4*S*,6*S*)-2-Methyl-6-nonylpiperidin-4-ol: (+)-Alkaloid **241D**. To a solution of **13** (0.135 g, 0.35 mmol 1 equiv) in MeOH (5 mL) was added Pd/C (5%, 15 mg) under H_2 atmosphere (1 atm). After 24 h, the solution was filtered through a pad of Celite and washed 3 times with MeOH. After concentration under vacuum, to the crude oil product was added slowly $NaBH_4$ (13 mg, 0.35 mmol, 1 equiv) at 0 °C. After 15 min at room temperature, the solution was quenched with a solution of brine and concentrated under vacuum. The residue was then diluted with ethyl acetate and washed with H_2O . Then the organic layer was dried and concentrated under vacuum. The yellow oil obtained was purified by flash chromatography (eluent, EtOAc to EtOAc/MeOH 90:10) to give (+)-alkaloid **241D** (71 mg, 84% over two steps) as colorless needles: mp 107–108 °C; $[\alpha]_D = +5.66$ (c 0.60

MeOH, 95% ee), lit.¹⁷ $[\alpha]_D = +5.90$ (c 0.65 MeOH, $\geq 99\%$ ee). 1H NMR (400 MHz, $CDCl_3$) δ 3.66 (tt, $J = 5.0, 11.2$ Hz, 1H), 2.69 (m, 1H), 2.55 (m, 1H), 1.96 (dt, $J = 11.2, 5.0$, 2H), 1.65–1.31 (m, 3H), 1.30–1.13 (m, 1H), 0.97 (q, $J = 11.2$ Hz, 1H), 0.91 (q, $J = 11.2$ Hz, 1H), 0.87 (t, $J = 7.0$ Hz, 1H).

■ ASSOCIATED CONTENT

Supporting Information

1H NMR and ^{13}C NMR of **5** to **11** are described and copies of spectra are given. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

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